

Object oriented programming

- Biopython is object-oriented
- Some knowledge helps understand how biopython works
- OOP is a way of organizing data and methods that work on them in a coherent package
- OOP helps structure and organize the code

Classes and objects

- A class:
 - is a user defined type
 - is a mold for creating objects
 - specifies how an object can contain and process data
 - represents an abstraction or a template for how an object of that class will behave
- An object is an instance of a class
- All objects have a type shows which class they were made from

Attributes and methods

- Classes specify two things:
 - attributes data holders
 - methods functions for this class
- Attributes are variables that will contain the data that each object will have
- Methods are functions that an object of that class will be able to perform

Class and object example

- Class: MySeq
- MySeq has:
 - attribute length
 - method translate
- An object of the class MySeq is created like this:
 - myseq = MySeq("ATGGCCG")
- Get sequence length:
 - myseq.length
- Get translation:
 - myseq.translate()

Summary

- An object has to be instantiated, i.e. created, to exist
- Every object has a certain type, i.e. is of a certain class
- The class decides which attributes and methods an object has
- Attributes and methods are accessed using . after the object variable name

Biopython

- Package that assists with processing biological data
- Consists of several modules some with common operations, some more specialized
- Website: biopython.org

Biopython must be installed

- Not part of python per se, has to be installed
- Several versions of python available, not all compatible with biopython
- On freebee, several python versions available:

[karinlag@freebee]~/teaching% which python
/usr/bin/python
[karinlag@freebee]~/teaching%

 module load python2 makes different version of python available:

[karinlag@freebee]~/teaching% module load python2
[karinlag@freebee]~/teaching% which python
/cluster/software/VERSIONS/python2-2.7.3/bin/python

Working with sequences

- Biopython has many ways of working with sequence data
- Covered today:
 - Alphabet
 - Seq
 - SeqRecord
 - SeqIO
- Other useful classes for working with alignments, blast searches and results etc are also available, not covered today

Class Alphabet

- Every sequence needs an alphabet
- CCTTGGCC DNA or protein?
- Biopython contains several alphabets
 - DNA
 - RNA
 - Protein
 - the three above with IUPAC codes
 - ...and others
- Can all be found in Bio.Alphabet package

Alphabet example

- Go to freebee
- Do module load python2 (necessary to find biopython modules) start python

NOTE: have to import Alphabets to use them

```
>>> import Bio.Alphabet
>>> Bio.Alphabet.ThreeLetterProtein.letters
['Ala', 'Asx', 'Cys', 'Asp', 'Glu', 'Phe', 'Gly', 'His', 'Ile',
'Lys', 'Leu', 'Met', 'Asn', 'Pro', 'Gln', 'Arg', 'Ser', 'Thr',
'Sec', 'Val', 'Trp', 'Xaa', 'Tyr', 'Glx']
>>> from Bio.Alphabet import IUPAC
>>> IUPAC.IUPACProtein.letters
'ACDEFGHIKLMNPQRSTVWY'
>>> IUPAC.unambiguous_dna.letters
'GATC'
>>>
```

Packages, modules and classes

- What happens here?
 - >>> from Bio.Alphabet import IUPAC
 >>> IUPAC.IUPACProtein.letters
- Bio and Alphabet are packages
 - packages contain other packages and modules
- IUPAC is a module
 - · a module is a file with python code
- IUPAC module contains class IUPACProtein and other classes specifying alphabets
- IUPACProtein has attribute letters

Seq

- Represents one sequence with its alphabet
- Methods:
 - translate()
 - transcribe()
 - complement()
 - reverse_complement()
 - • •

Using Seq

```
Import classes
>>> from Bio.Seg import Seg
                                Create object
>>> import Bio.Alphabet
>>> seg = Seg("CCGGGTT", Bio.Alphabet.IUPAC.unambiguous dna)
>>> seq
Seg('CCGGGTT', IUPACUnambiguousDNA())
>>> seg.transcribe()
Seg('CCGGGUU', IUPACUnambiquousRNA())
                                       Use methods
>>> seq.translate()
Seg('PG', IUPACProtein())
>>> seg = Seg("CCGGGUU", Bio.Alphabet.IUPAC.unambiguous rna)
                                             New object, different alphabet
>>> seq.transcribe()
Traceback (most recent call last):
  File "<stdin>", line 1, in <module>
 File "/site/VERSIONS/python-2.6.2/lib/python2.6/site-packages/Bio/Seg.py",
 line 830, in transcribe
    raise ValueError("RNA cannot be transcribed!")
ValueError: RNA cannot be transcribed!
>>> seq.translate()
                                       Alphabet dictates which
Seq('PG', IUPACProtein())
                                       methods make sense
>>>
```

Seq as a string

- Most string methods work on Seqs
- If string is needed, do str(seq)

```
>>> seq = Seq('CCGGGTTAACGTA', Bio.Alphabet.IUPAC.unambiguous dna)
>>> seq[:5]
Seq('CCGGG', IUPACUnambiguousDNA())
>>> len(seg)
13
>>> seq.lower()
Seq('ccgggttaacgta', DNAAlphabet())
>>> print seg
CCGGGTTAACGTA
>>> mystring = str(seq)
>>> print mystring
CCGGGTTAACGTA
>>> type(seq)
<class 'Bio.Seq.Seq'>
>>> type(mystring)
                             How to check what class
<type 'str'>
                             or type an object is from
>>>
```

SeqRecord

- Seq contains the sequence and alphabet
- But sequences often come with a lot more
- SeqRecord = Seq + metadata
- Main attributes:
 - id name or identifier
 - seq seq object containing the sequence

```
Existing sequence
```

```
>>> seq
Seq('CCGGGTTAACGTA', IUPACUnambiguousDNA()) SeqRecord is a class
>>> from Bio.SeqRecord import SeqRecord
>>> seqRecord = SeqRecord(seq, id='001')
>>> seqRecord
SeqRecord(seq=Seq('CCGGGTTAACGTA', IUPACUnambiguousDNA()),
id='001', name='<unknown name>', description='<unknown description>',
dbxrefs=[])
>>>
```

SeqRecord attributes

From the biopython webpages:

Main attributes:

id - Identifier such as a locus tag (string)seq - The sequence itself (Seq object or similar)

Additional attributes:

name - Sequence name, e.g. gene name (string)
 description - Additional text (string)
 dbxrefs - List of database cross references (list of strings)
 features - Any (sub)features defined (list of SeqFeature objects)
 annotations - Further information about the whole sequence (dictionary)
 Most entries are strings, or lists of strings.

letter_annotations - Per letter/symbol annotation (restricted dictionary). This holds Python sequences (lists, strings or tuples) whose length matches that of the sequence. A typical use would be to hold a list of integers representing sequencing quality scores, or a string representing the secondary structure.

SeqRecords in practice...

```
>>> from Bio.SegRecord import SegRecord
>>> from Bio.Seg import Seg
                                             Import necessary classes
>>> from Bio.Alphabet import DNAAlphabet
>>> segRecord = SegRecord(Seg('GCAGCCTCAAACCCCAGCTG',
... DNAAlphabet), id = 'NM 005368.2', name = 'NM 005368',
... description = 'Myoglobin var 1',
                                                Create object
... dbxrefs = ['GeneID:4151', 'HGNC:6915'])
>>>
>>> seqRecord
                                                Print object
SeqRecord(seq=Seq('GCAGCCTCAAACCCCAGCTG',
 <class 'Bio.Alphabet.DNAAlphabet'>), id='NM 005368.2',
name='NM 005368', description='Myoglobin var 1',
dbxrefs=['GeneID:4151', 'HGNC:6915'])
>>>
```

SeqIO

- How to get sequences in and out of files
- Retrieves sequences as SeqRecords, can write SeqRecords to files
- Reading:
 - SeqIO.parse(filehandle, format)
 - returns a generator that gives SeqRecords
- Writing:
 - SeqIO.write(SeqRecord(s), filehandle, format)

NOTE: examples in this section from http://biopython.org/wiki/SeqIO

SeqIO formats

- List: http://biopython.org/wiki/SeqIO
- Some examples:
 - fasta
 - genbank
 - several fastq-formats
 - ace
- Note: a format might be readable but not writable depending on biopython version

Reading a file

```
from Bio import SeqIO
fh = open("example.fasta", "r")
for record in SeqIO.parse(fh,"fasta"):
    print record.id
fh.close()
```

- SeqIO.parse returns a SeqRecord iterator
- An iterator will give you the next element the next time it is called
- Useful because if a file contains many records, we avoid putting all into memory all at once

Parsing fasta files

- Copy fasta file containing 3 sequences cp ~karinlag/teaching/mb.fsa .
- In python interactive shell:

XM 001081975.2

NM 001164047.1

CCTCTCCCCA

TAGCTGCCCA

>>>

- Goal: convert from genbank to fasta
- cp ~karinlag/teaching/mb.gbk .
- Create script file:
 - Import both sys and SeqIO
 - Take in file name as sys.argv[1]
 - For each record in file (remember:genbank!)
 - Print record
 - Close file
 - Save as convert.py
- Run script with mb.gbk

- Modification 1:
 - Print
 - The id
 - The description
 - The sequence

```
[karinlag@freebee]~/teaching% cat convert1.py
from Bio import SeqIO
import sys

# Open the input file
fh = open(sys.argv[1], "r")

for record in SeqIO.parse(fh, "genbank"):
    # Print only id, description and sequence
    print record.id
    print record.description
    print record.seq
fh.close()
[karinlag@freebee]~/teaching%
```

Modifications

- Figure out how to:
 - print the description of each genbank entry
 - which annotations each entry has
 - print the taxonomy for each entry
- Description:
 - seqRecord.description
- Annotations:
 - seqRecord.annotations.keys()
- Taxonomy:
 - seqRecord.annotations['taxonomy']

Writing files

```
from Bio import SeqIO
sequences = ... # add code here
output_handle = open("example.fasta", "w")
SeqIO.write(sequences, output_handle, "fasta")
output_handle.close()
```

- Note: sequences is here a list containing several SeqRecords
- Can write any iterable containing SeqRecords to a file
- Can also write a single sequence

- Modification 2:
 - Get output file name as sys.argv[2]
 - Open outfile
 - Per record,
 - Write it to file in fasta format
 - Close input file
 - Close output file
- Bonus question: can you think of how you would add the organism name to the id?

```
[karinlag@freebee]~/teaching% cat convert2.py
from Bio import SeqIO
import sys
# Open the input file
                                     Open both input
fh = open(sys.argv[1], "r")
                                     and output file
# Open the output file
fo = open(sys.argv[2],
for record in SeqIO.parse(fh, "qenbank"):
    # Use SeqIO to write properly
    # formatted record
    SeqIO.write(record, fo, "fasta")
                                         Write out record
fh.close()
fo.close()
# ...and closing files
[karinlag@freebee]~/teaching%
```

Bonus question

- How to add organism name:
 - get the taxonomy list from the annotation dictionary
 - get the last element of the list with slicing off
 -1, the last element
 - concat the record.id with the orgname

```
orgname = record.annotations['taxonomy'][-1]
print record.id + "_" + orgname
```

Tips and hints

- Always comment your code easier to understand later
- Never write lots of code without testing while writing – makes for less code to debug
- Always test on input where you know what the results should be
- If it went to easy, too well or too fast: it is probably wrong!

Learning more

- Recommended book:
 - Sebastian Bassi:Python for Bioinformatics
- www.python.org
 - has lots of documentations and beginner tutorials
- Google

