

Bioinformatics for molecular biology

Structural bioinformatics tools, predictors, and 3D modeling – Structural Biology Review

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(<http://www.ous-research.no/rognes>)

CF: Bioinformatics services

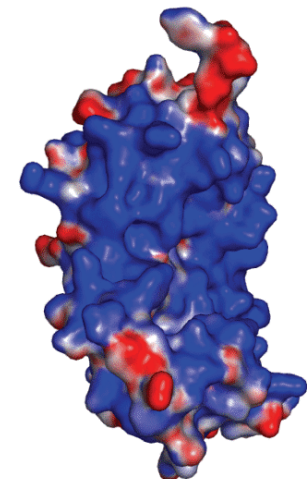
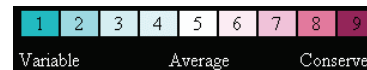
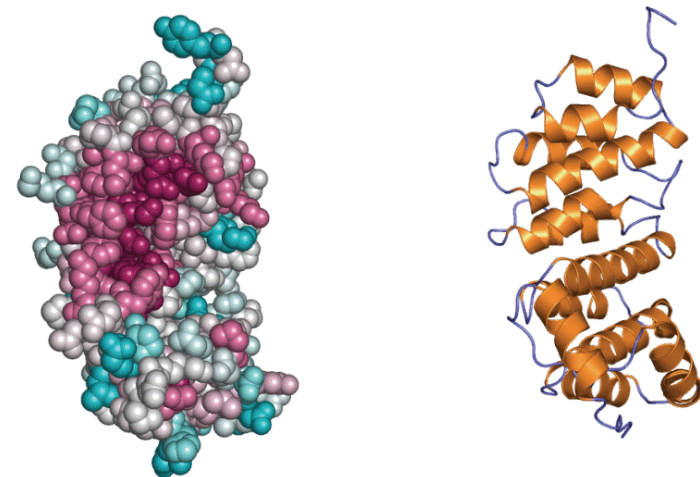
(<http://core.r-r-research.no/bioinformatics>)

CLS: Bioinformatics education

(<http://www.mn.uio.no/ifi/english/research/networks/clsi>)

Main research area:

Structural and Applied Bioinformatics



Overview

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Structural Bioinformatics

Now:

- Protein Structure Review
 - Amino acids, polypeptides, secondary structure elements, visualization, structure determination by X-ray crystallography and NMR methods, PDB

Later...

- Structure comparison and classification (CASP & SCOP)
- Predictors
- 3D structure modeling
 - *Ab initio*
 - Threading/fold recognition
 - Homology modeling
- Practical exercises
 - PyMOL & visualization
- Practical Exercises
 - Homology modeling of influenza neuraminidase (Tamiflu resistance?)
 - Other homology modeling
 - Threading
 - Your own project?



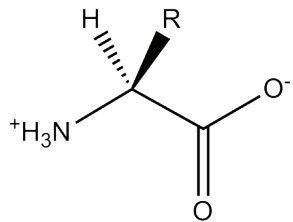
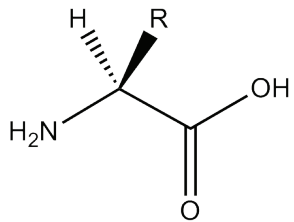
Stop me and ask questions!!

Amino acids – the building blocks of proteins

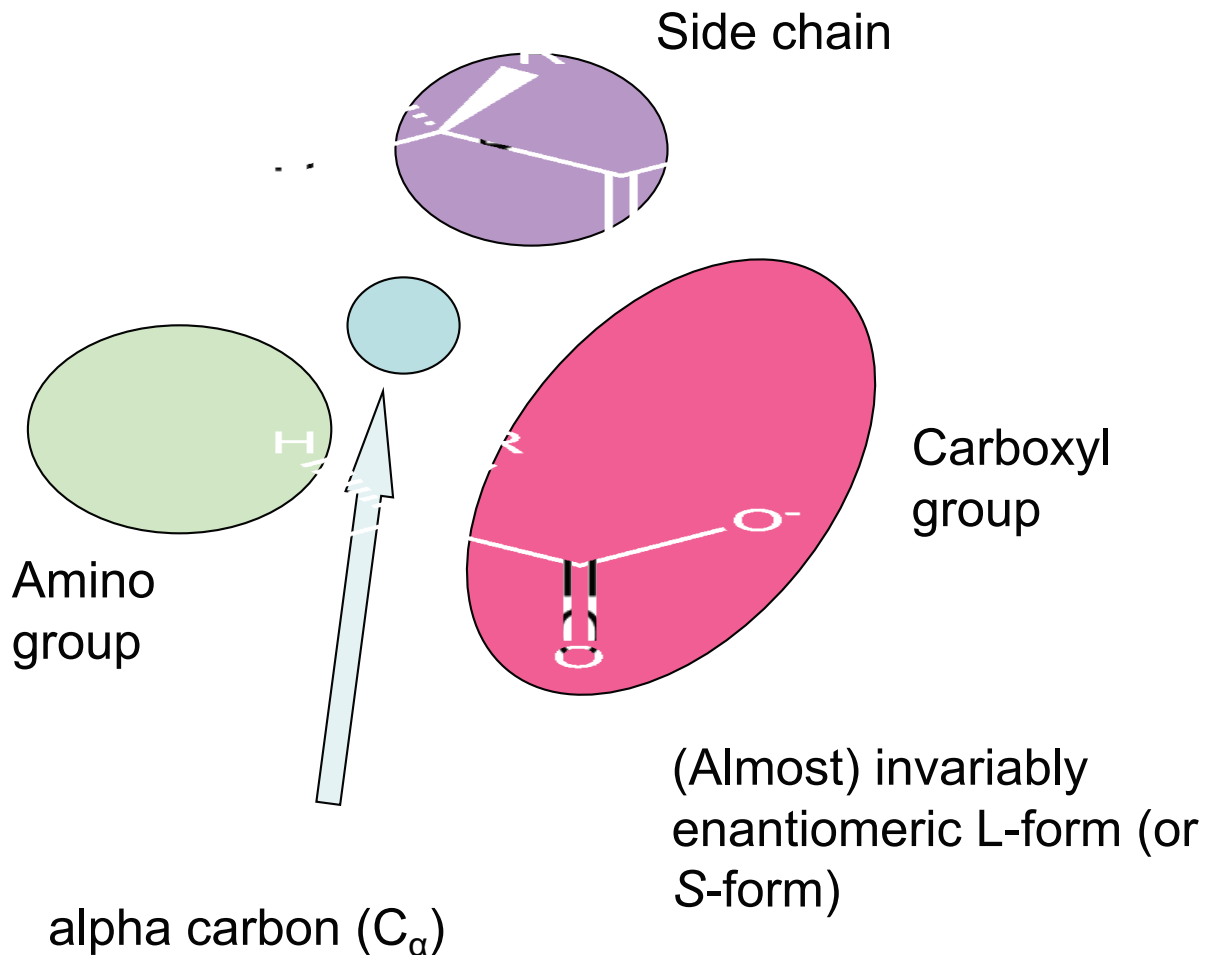
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Structural Bioinformatics

Proteins are built from 20 naturally occurring amino acids. They have an amino (-NH_2) and acidic (-COOH) functional group

The side chain group (R) determines the properties of the amino acid

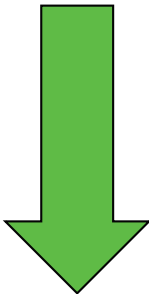


Zwitterionic form
found at
physiological pH



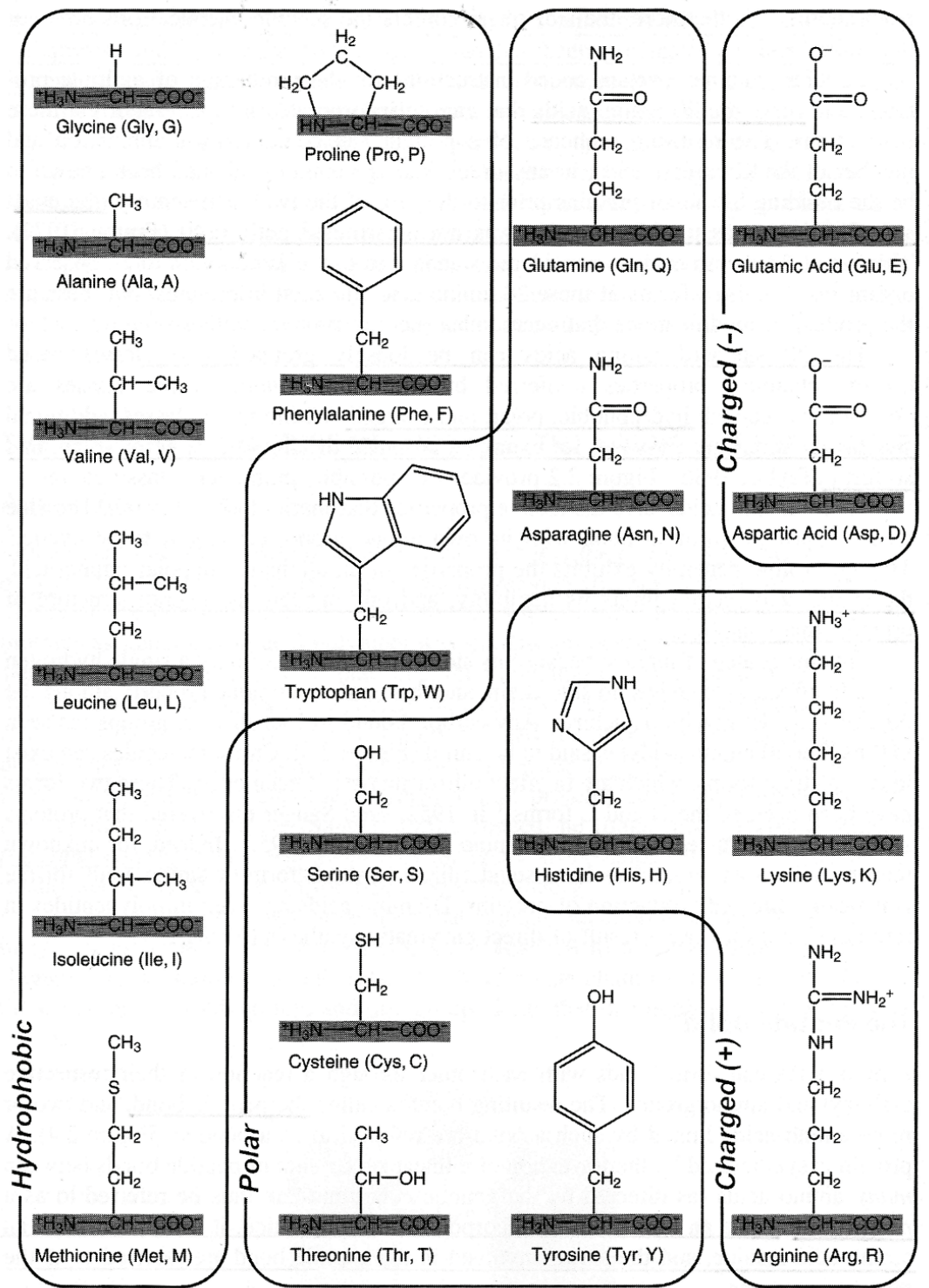
Amino acids

R-group properties:

- Large
 - Small
 - Hydrophobic
 - Aliphatic
 - Aromatic
 - Polar
 - Charged
 - Positive/negative charge
- 

Increasing hydrophilicity/higher water (solvent) affinity

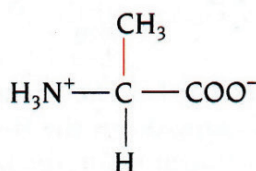
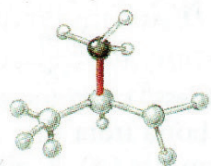
Structural Bioinformatics,
Eds. P.E. Bourne & H. Weissig
(Wiley, Hoboken, NJ, 2003)



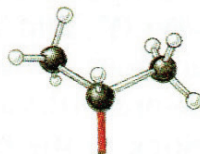
Amino acids

Introduction to Protein Structure, C. Branden & J. Tooze
(Garland, New York, 1998)

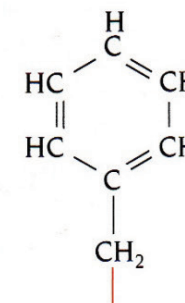
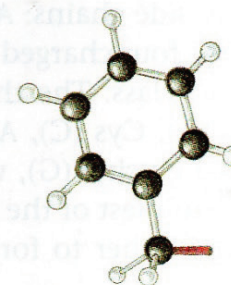
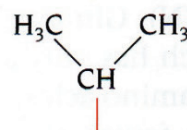
(a) Hydrophobic amino acids



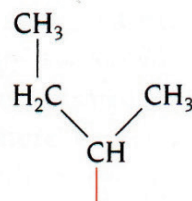
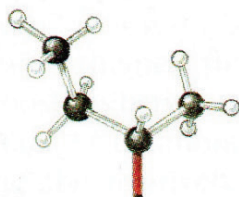
A Ala, Alanine



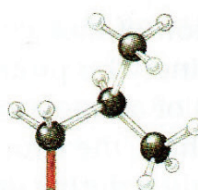
V Val, Valine



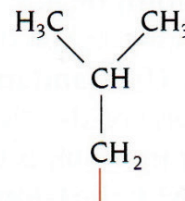
F Phe, Phenylalanine



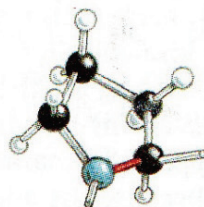
I Ile, Isoleucine



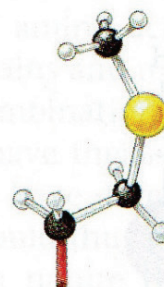
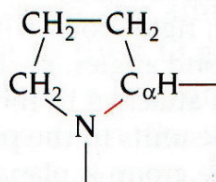
L Leu, Leucine



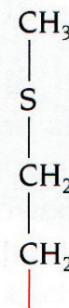
- Hydrophobic
- Aliphatic
- Aromatic



P Pro, Proline



M Met, Methionine

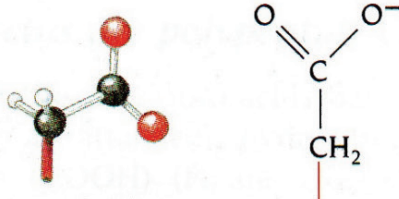


- 3-letter code
- 1-letter code

Amino acids

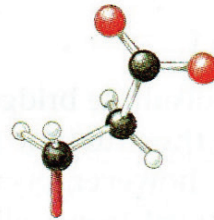
Introduction to Protein Structure, C. Branden & J. Tooze
(Garland, New York, 1998)

(b) Charged amino acids



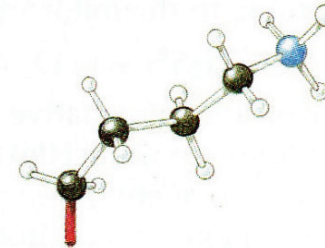
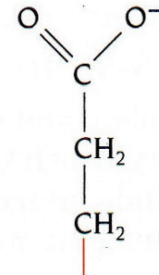
D Asp, Aspartic acid

Aspartate

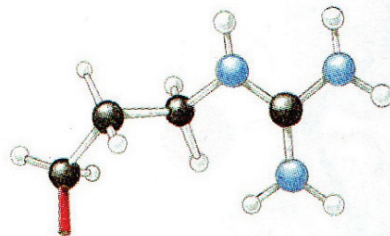
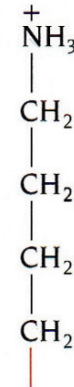


E Glu, Glutamic acid

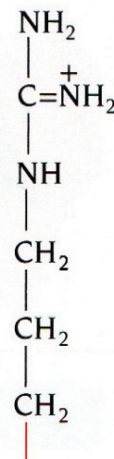
Glutamate



K Lys, Lysine



R Arg, Arginine



- Hydrophilic
 - Positive charge/basic
 - Negative charge/acidic

(d) Glycine



G Gly, Glycine

Amino acids

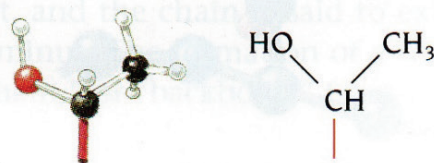
Introduction to Protein Structure, C. Branden & J. Tooze
(Garland, New York, 1998)

(c) Polar amino acids

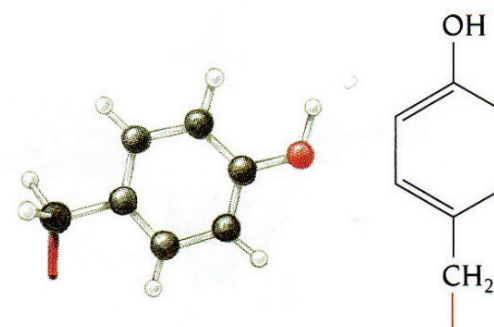
• Hydrophilic



S Ser, Serine



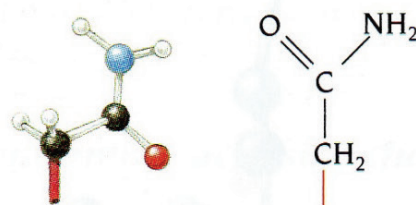
T Thr, Threonine



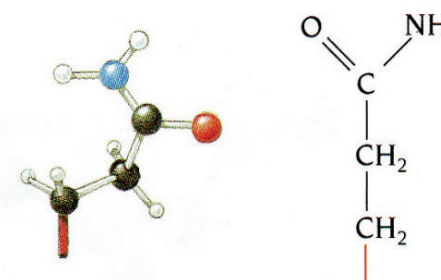
Y Tyr, Tyrosine



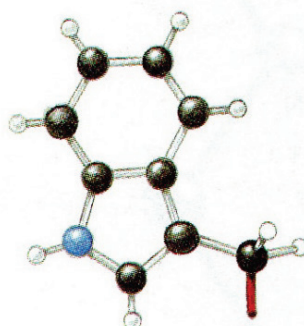
C Cys, Cysteine



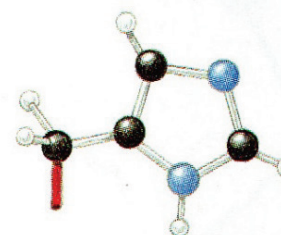
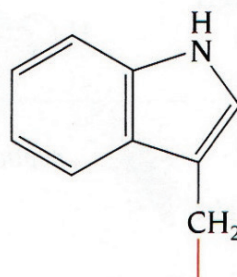
N Asn, Asparagine



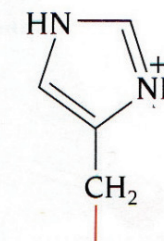
Q Gln, Glutamine



W Trp, Tryptophan

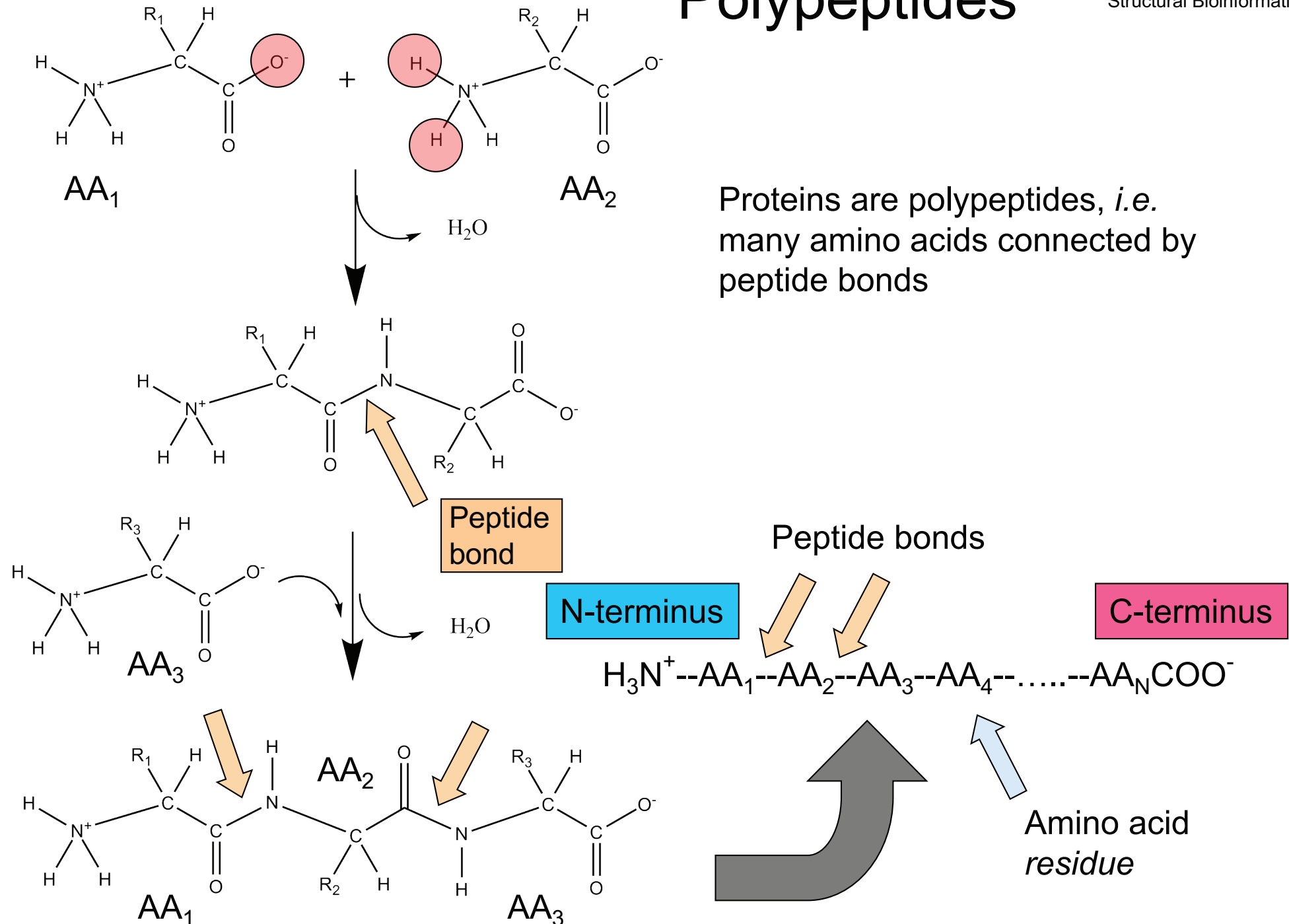


H His, Histidine



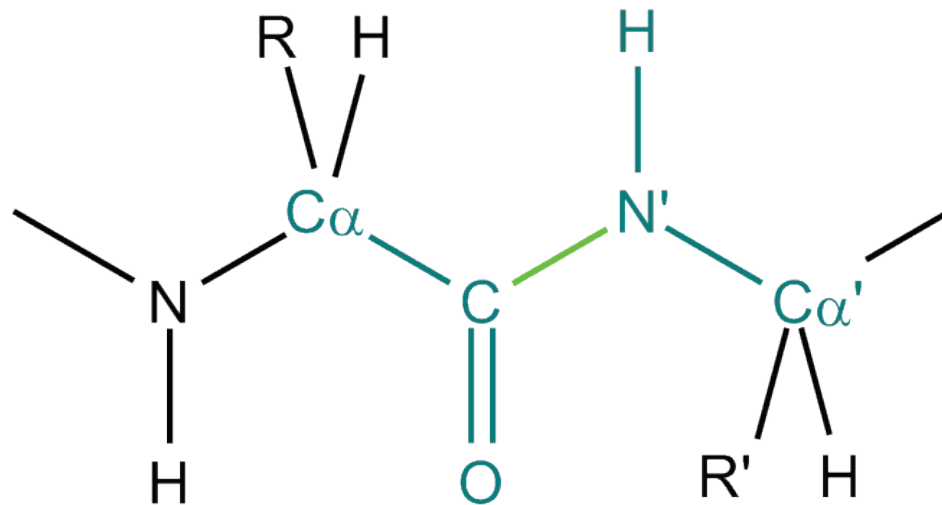
Polypeptides

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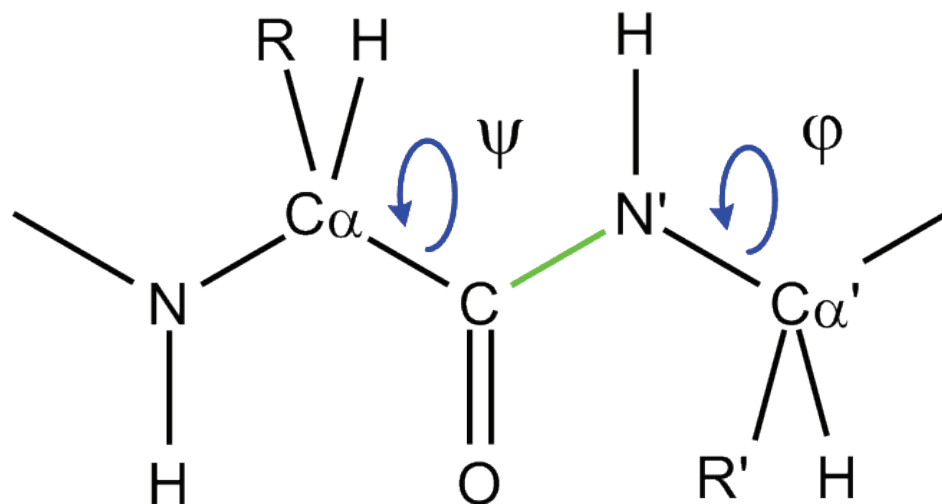
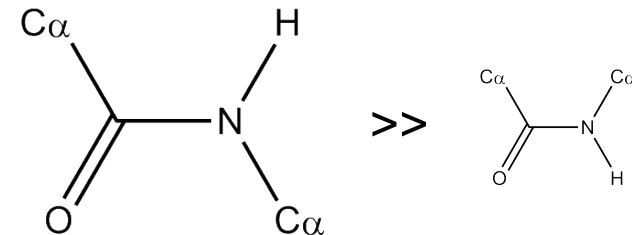
Dihedral angles

Proteins are polypeptides, *i.e.* many amino acids connected by peptide bonds



The peptide bond (light green) is a partial double bond and is fixed at $\sim 180^\circ$, *i.e.* the green part is flat

Cis-form for peptide bond is extremely rare except for prolines ($\sim 25\%$).

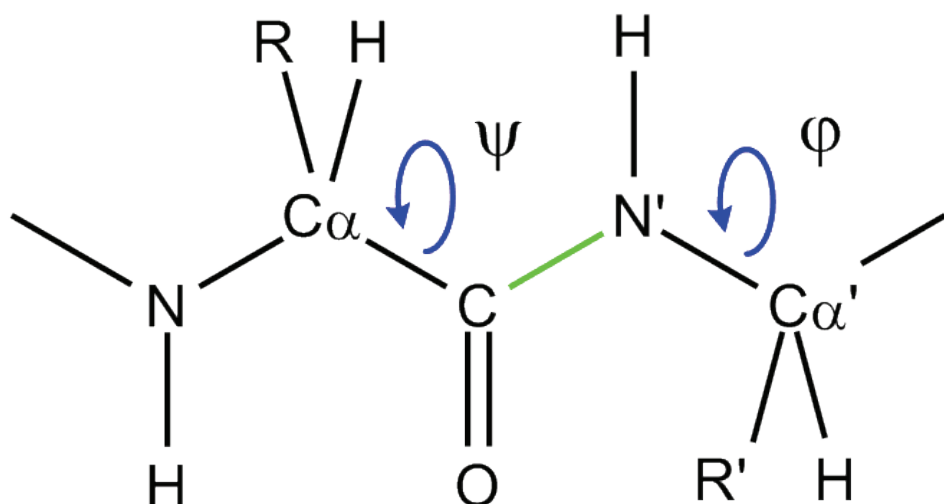


The dihedral angles phi (ϕ) and psi (ψ) determines the conformation of the peptide backbone

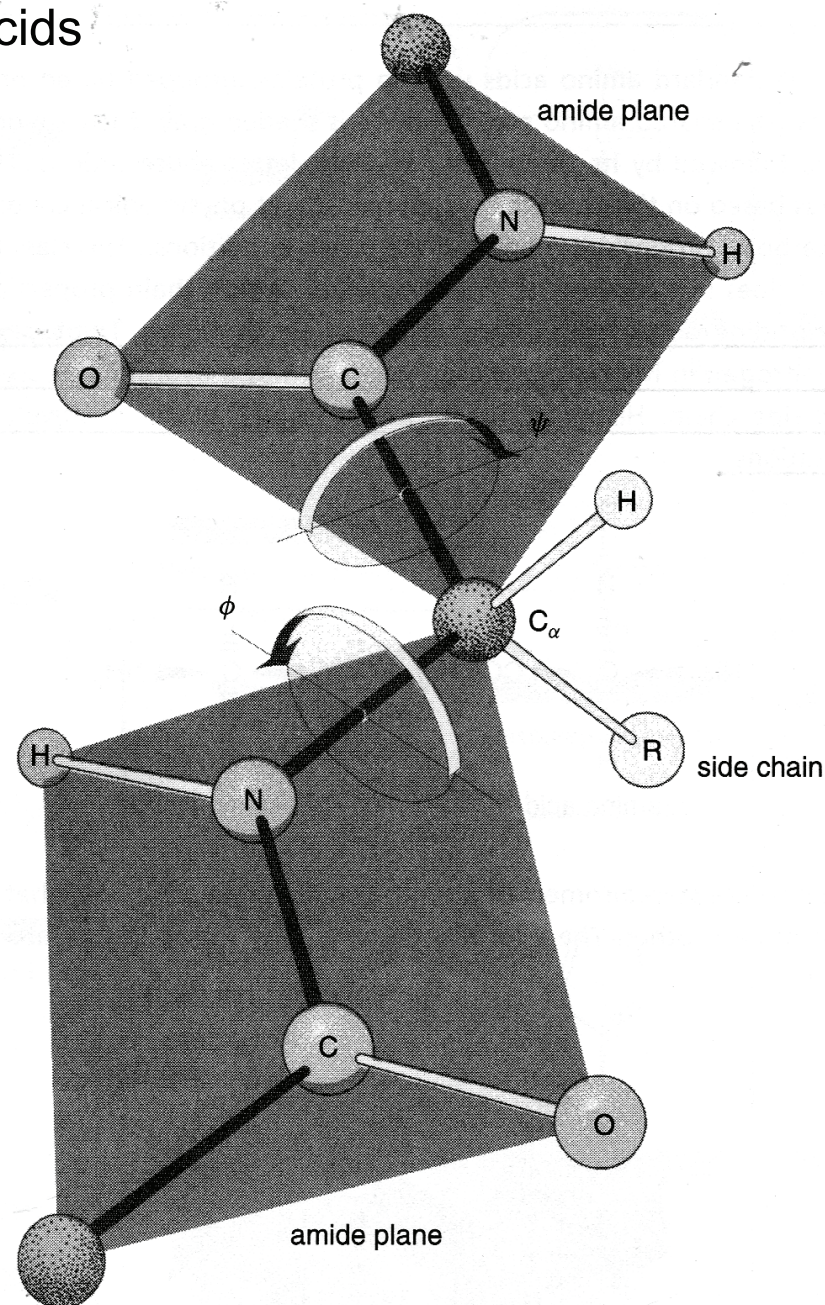
Dihedral angles

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Proteins are polypeptides, *i.e.* many amino acids connected by peptide bonds



One (ϕ, ψ) pair for each residue in a protein

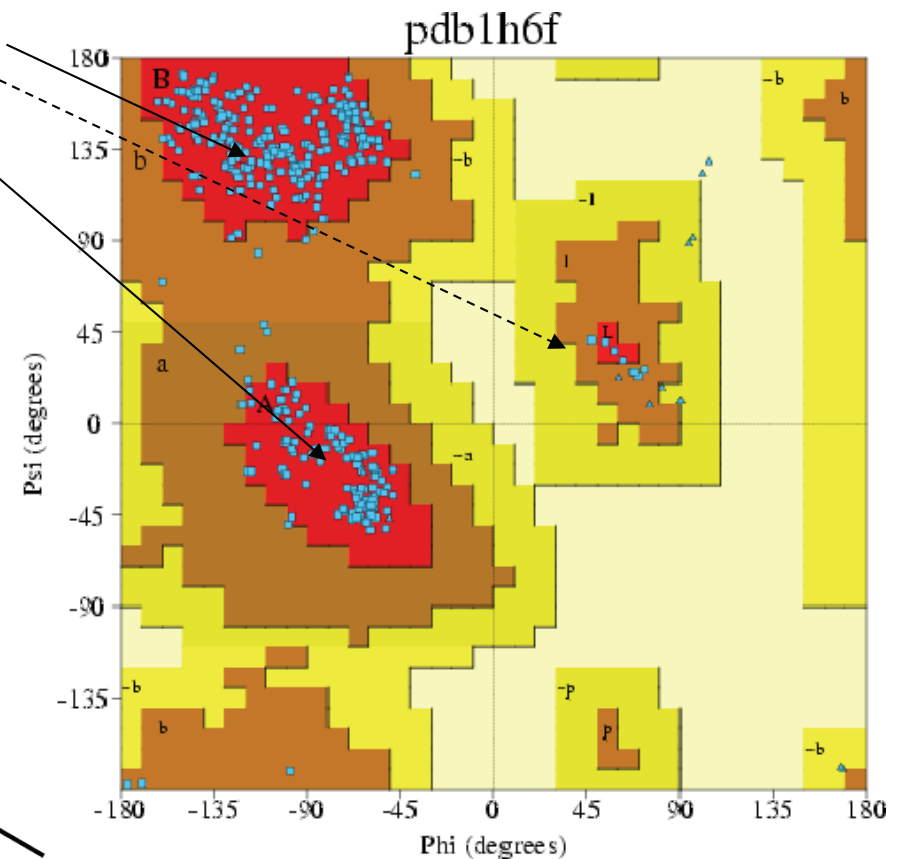
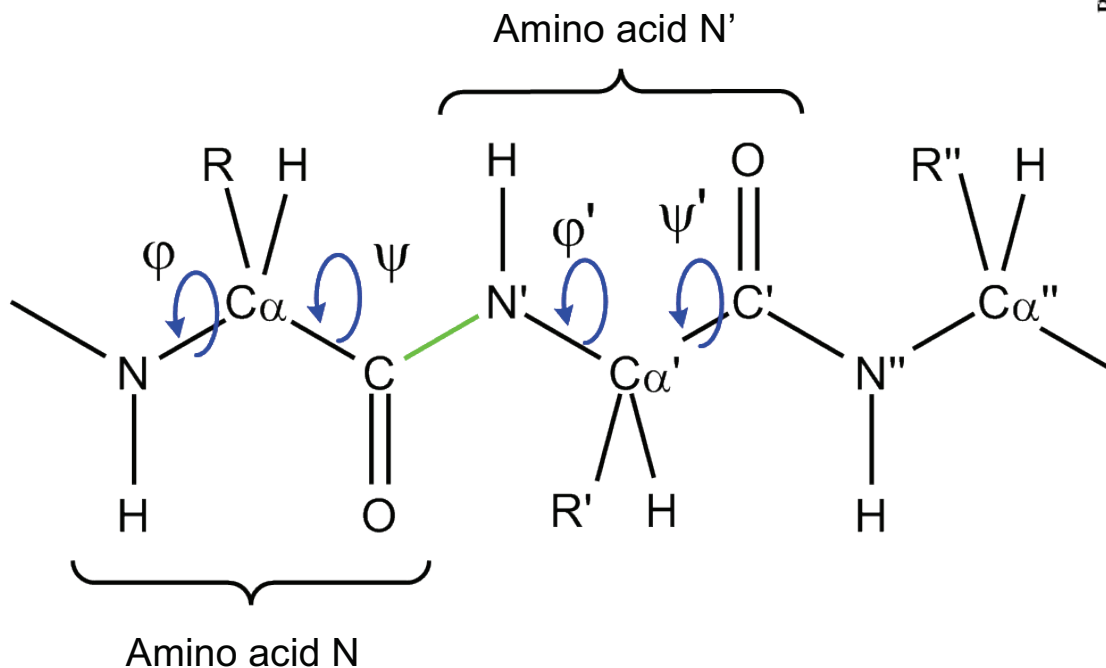


Structural Bioinformatics,
Eds. P.E. Bourne & H. Weissig
(Wiley, Hoboken, NJ, 2003)

Ramachandran plot

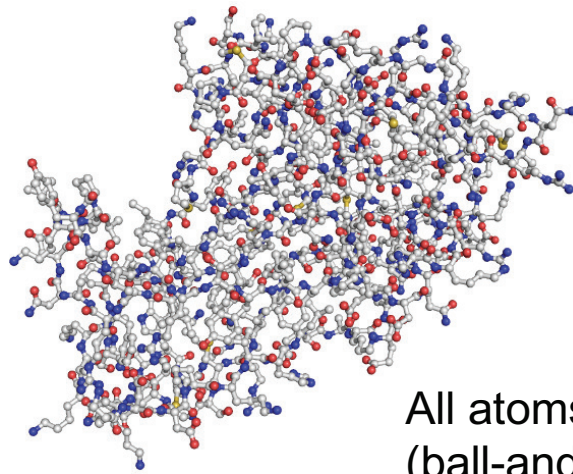
- Dihedral angles
 - Phi (ϕ)
 - Psi (ψ)
- Plot of (ϕ, ψ) angle pairs for each residue in a protein:
Ramachandran plot

Most (ϕ, ψ) pairs in
two (three) regions

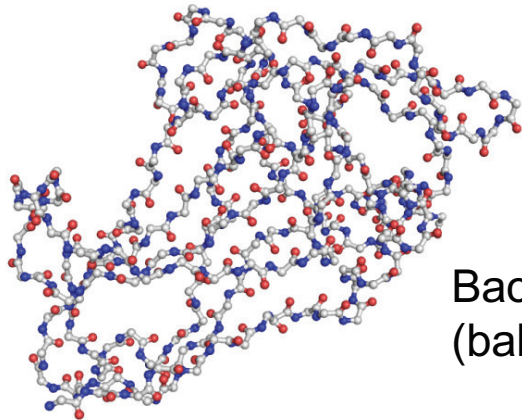


One point (blue spot) for each of
the 184 residues in this protein
(1H6F) (a human a transcription
factor)

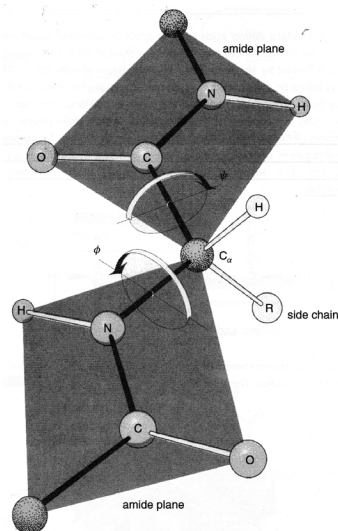
Ramachandran plot



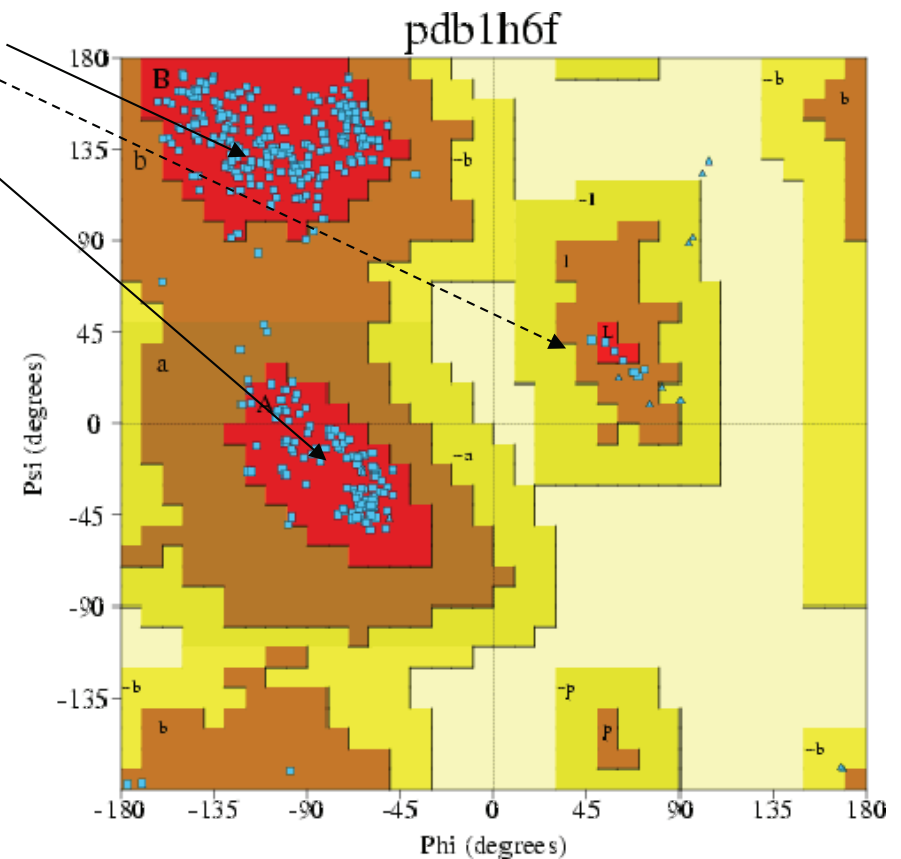
All atoms
(ball-and-stick)



Backbone atoms only
(ball-and-stick)



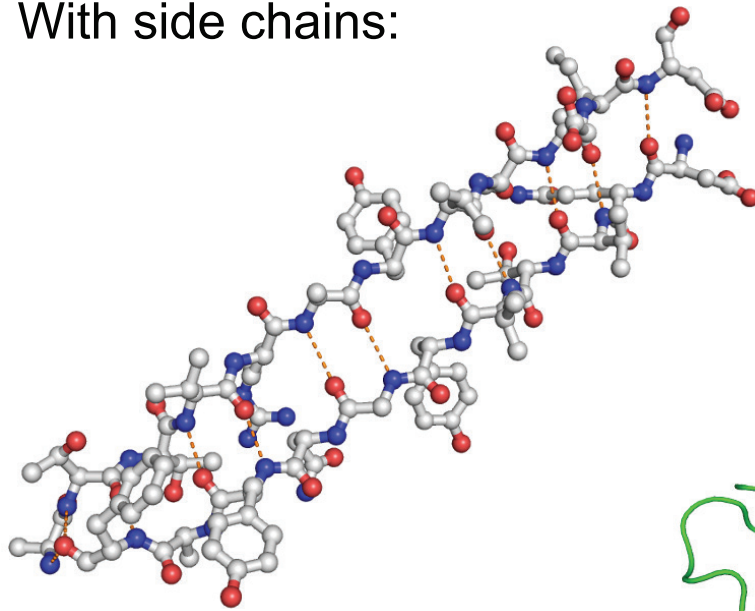
Most (ϕ, ψ) pairs in
two (three) regions



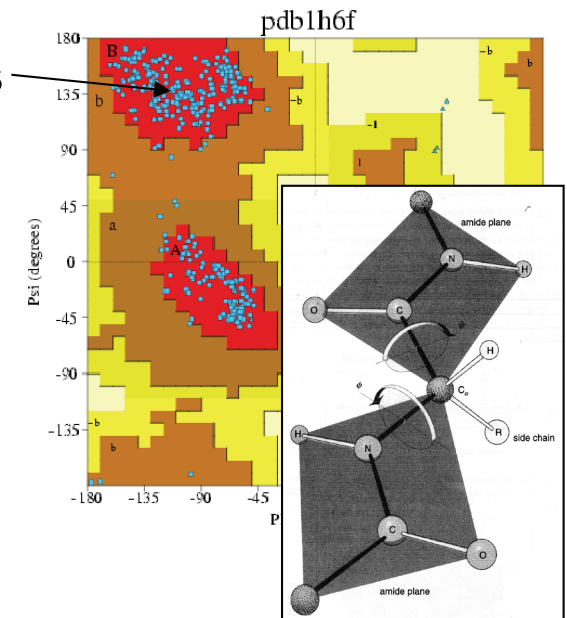
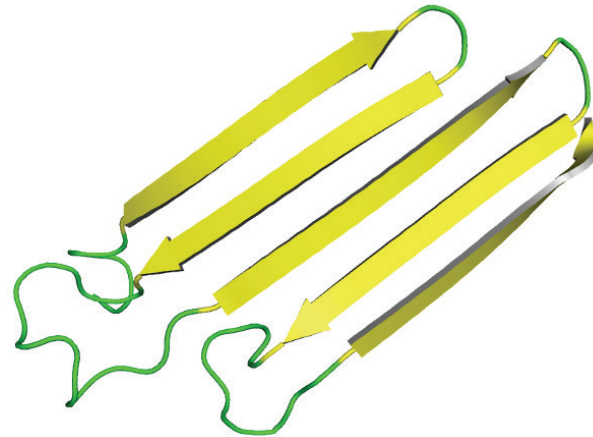
One point (blue spot) for each of
the 184 residues in this protein
(1H6F) (a human α transcription
factor)

Secondary structure – β -sheets

With side chains:

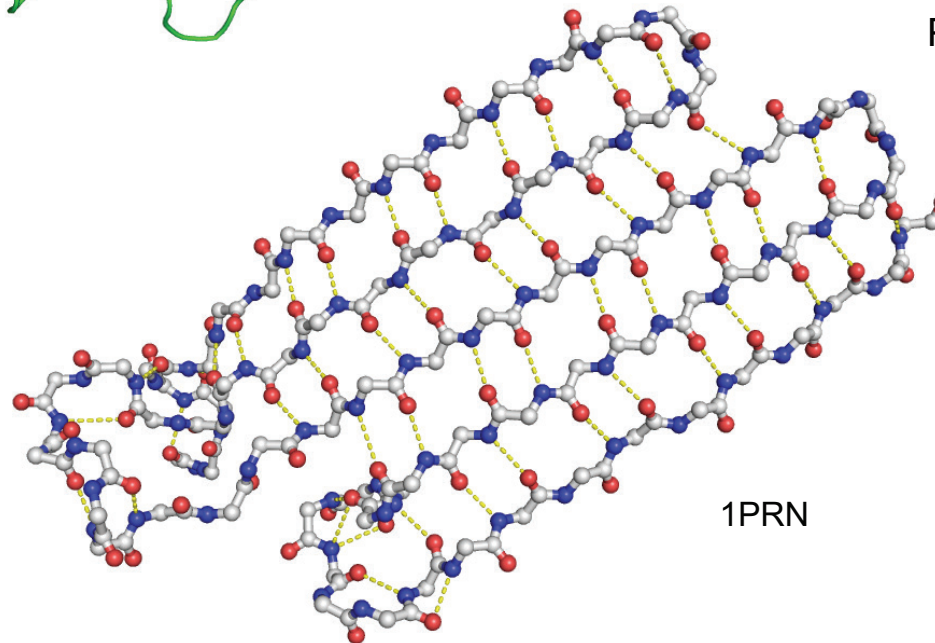
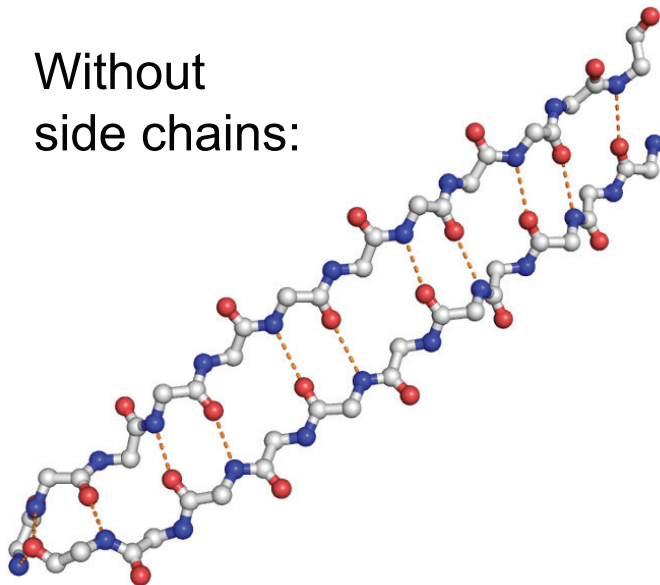


β -strands & β -sheets



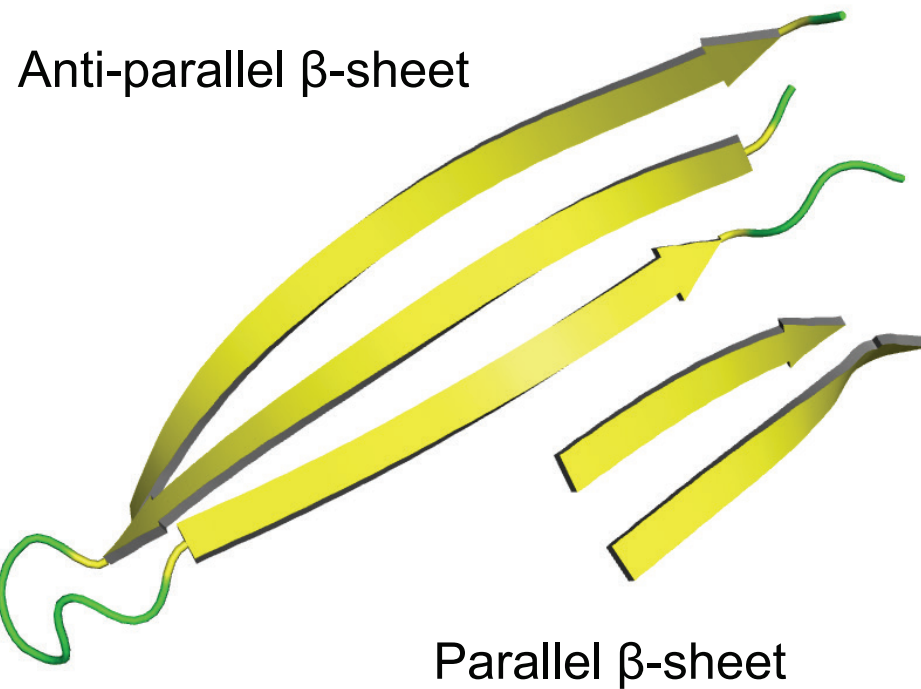
Psi ~ 135°
Phi ~ -100°

Without
side chains:

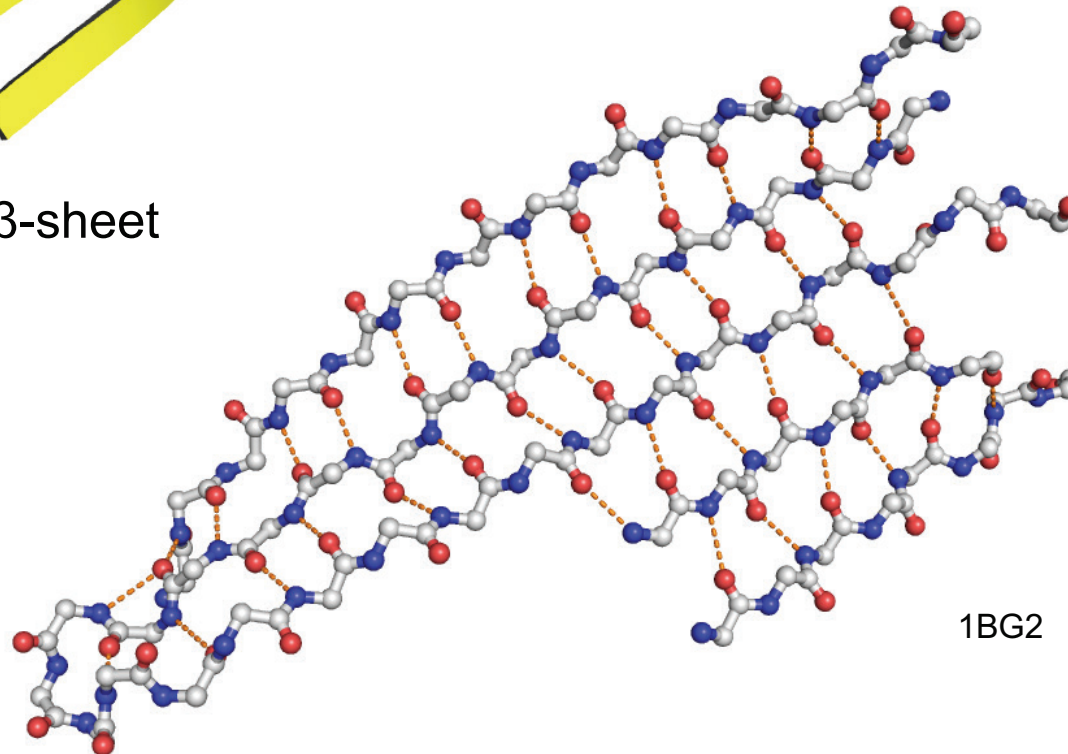


1PRN

Secondary structure – β -sheets

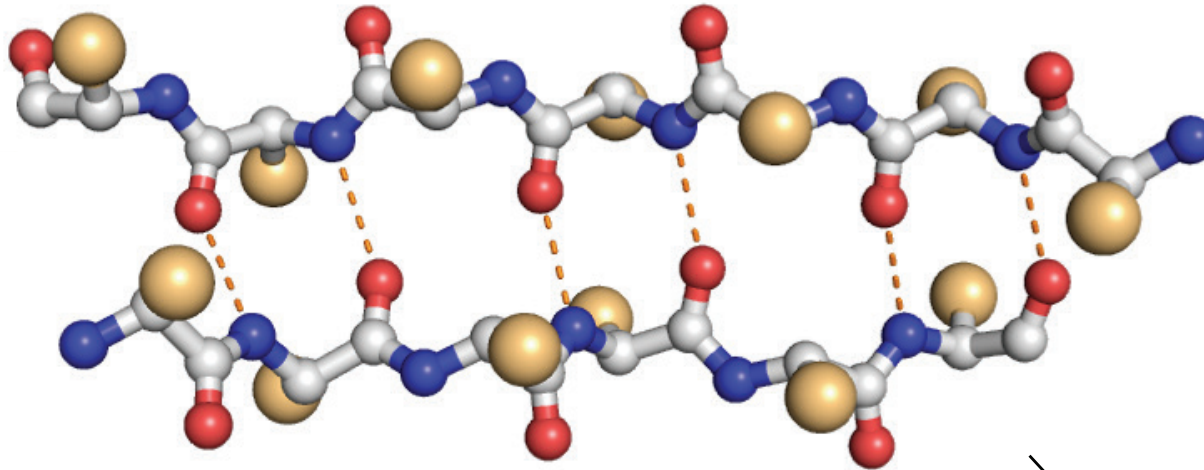


β -sheets can be
parallel, anti-
parallel or mixed

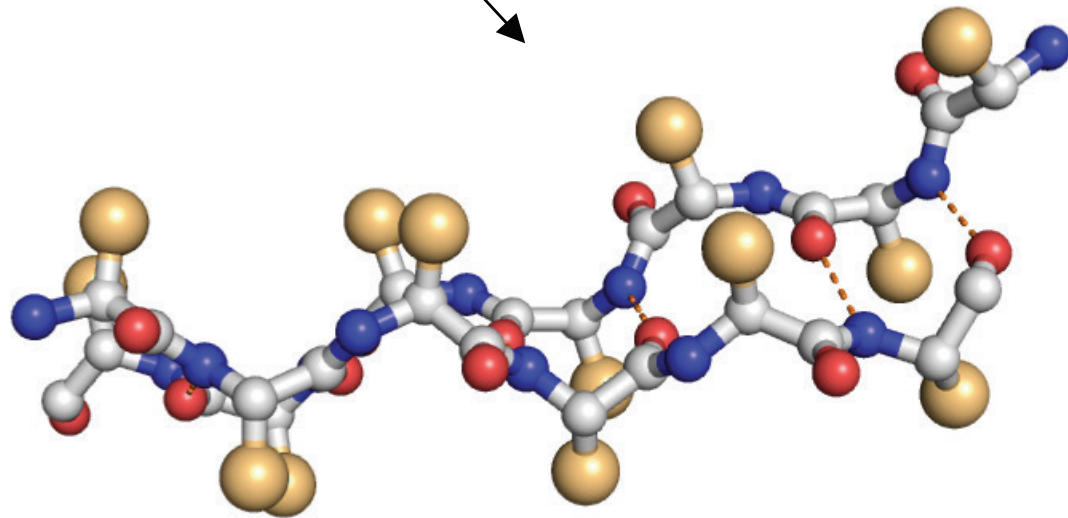


1BG2

Secondary structure – β -sheets



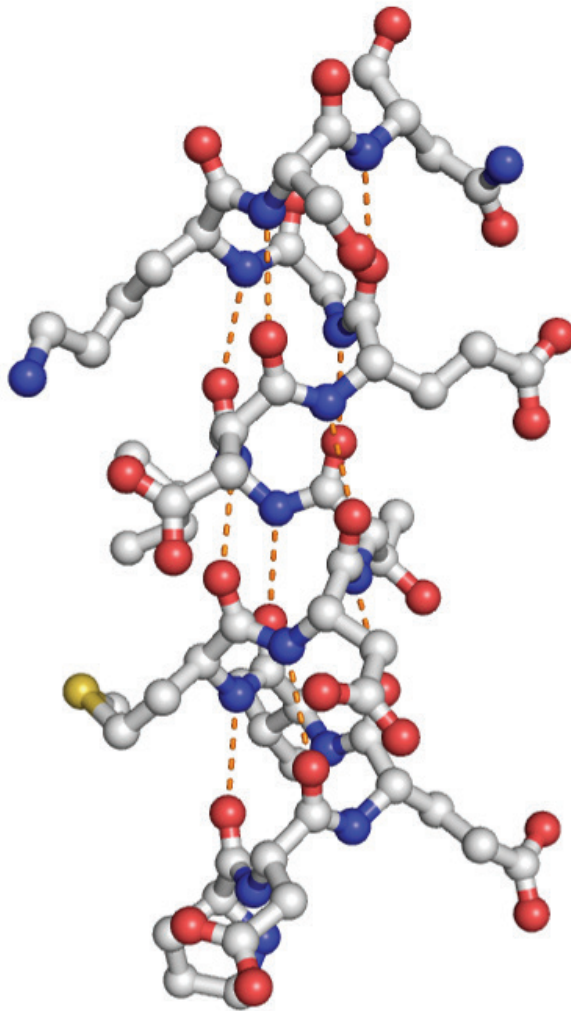
90° rotation



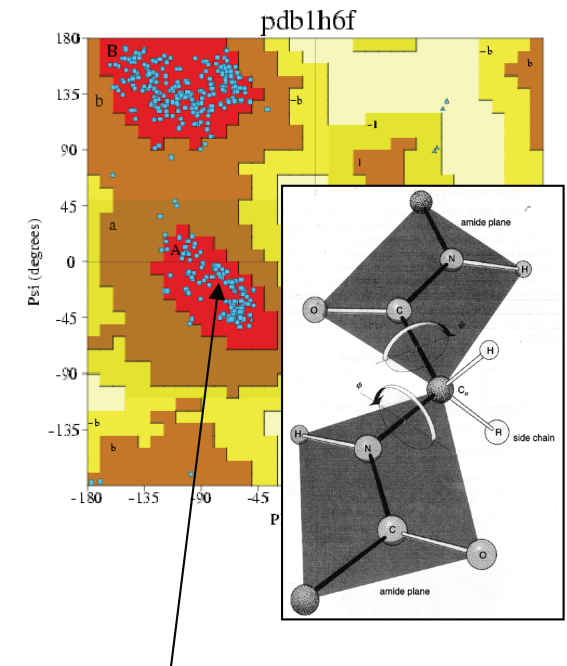
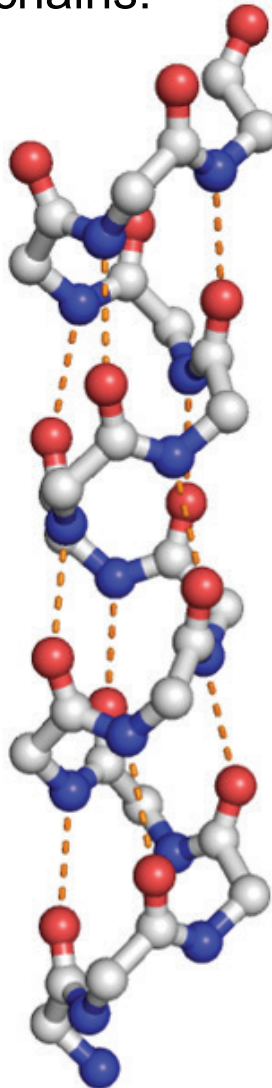
In β -sheets each side chain R-group is alternately on opposite sides of the plane of the sheet

Secondary structure – α -helices

With side chains:



Without side chains:



α -helix

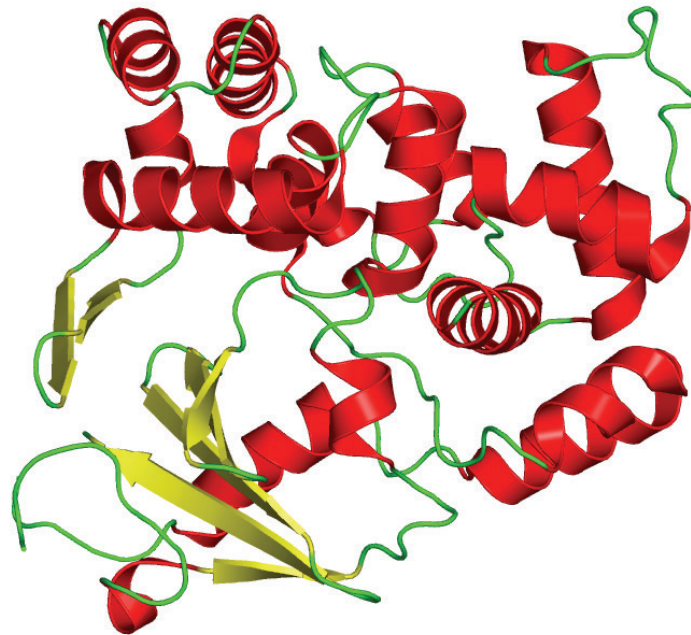
3.6 amino acids/turn

H-bonds between amino acids n & $n+4$

Partial positive charge at N-terminus and negative charge at C-terminus, *i.e.* it is a *dipole*

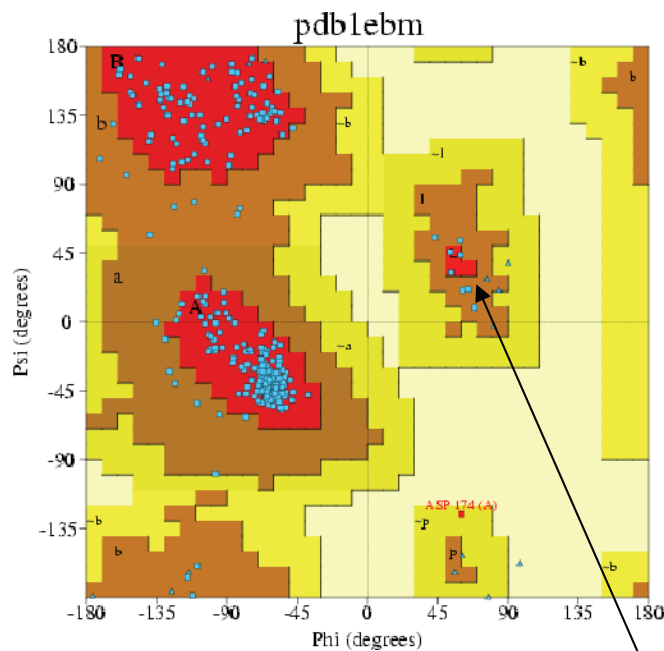
Secondary structure – 3 states

Three "states":
 α -helices (H)
 β -sheets (E)
Loops/coils (C)



Loops/coils:

- Loops may be hairpins or sharp turns
- Random coils/irregular loops
- Often "allowed" with insertions/deletions, *i.e.* evolutionary variable regions



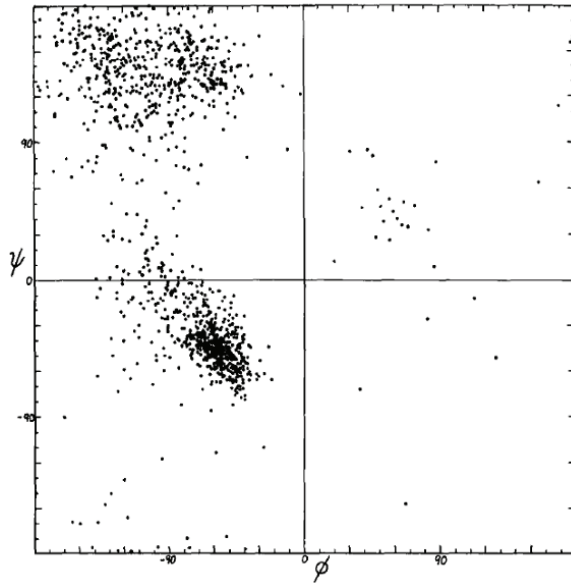
Left-handed helices

Coil here: "Everything that is not helix or sheet"

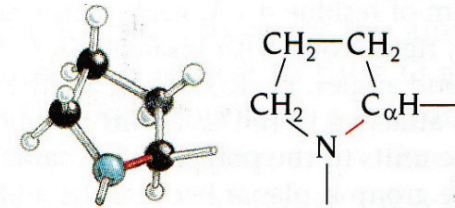
Coil often means: "Everything that is not helix or sheet or some characteristic loops"

Often contains Gly (to give flexibility) or Pro (to "break up" secondary structure elements)

Secondary structure – Gly & Pro

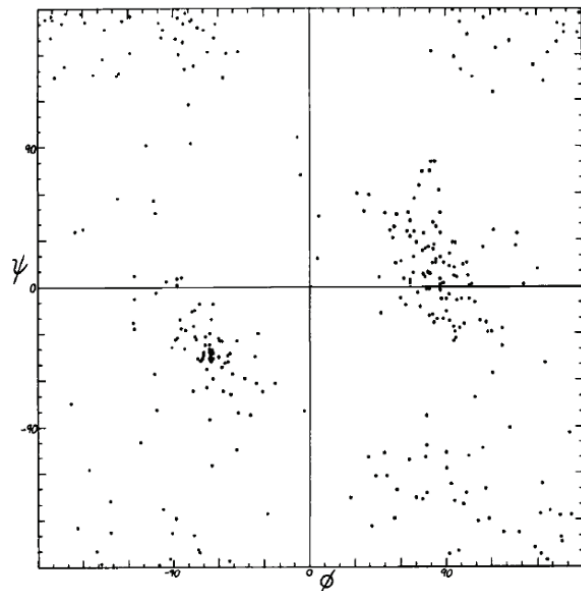


Non-glycine residues are mainly in α -helices and β -sheets



P Pro, Proline

Proline has very little flexibility in the backbone (disruptive to normal secondary structure)



Glycine has no side chain and a more flexible backbone

(d) Glycine



G Gly, Glycine

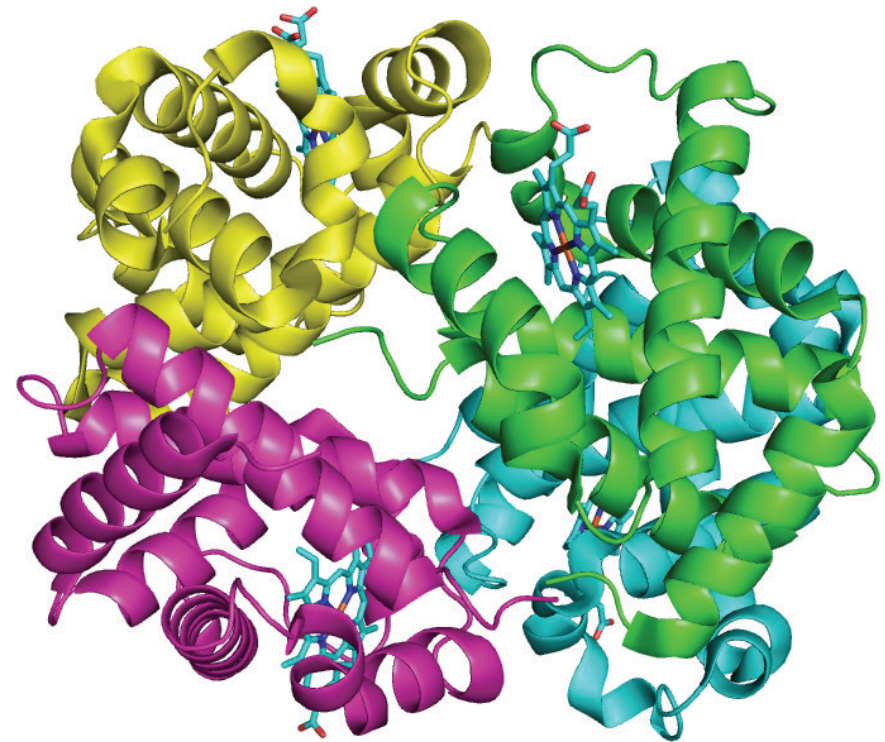
Protein structure

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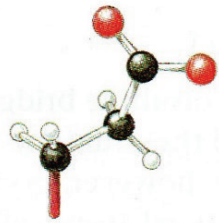
- Primary structure: Linear amino acid sequence
- Secondary structure: Local conformation of the peptide chain:
 - α -helix
 - β -sheet
- Tertiary structure: The full 3D structure
- Quaternary structure: Association of several proteins/peptide chains into protein complexes

Met-Ala-Leu-Asp-Asp-...

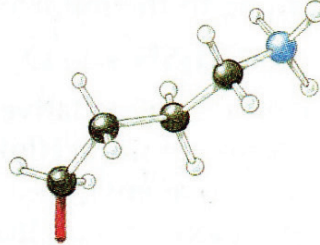
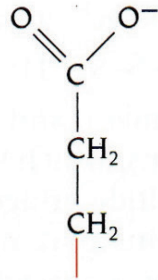
Hemoglobin, 1GZX



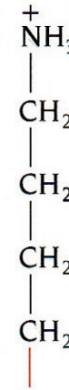
Residue properties



E Glu, Glutamic acid



K Lys, Lysine



pK_a depends on local environment

e.g. Glu close to negatively charged moiety: higher pK_a
Glu close to Lys is more willing to give off H^+ , i.e. lower pK_a

$pK_a = 4.25$

$pK_a = 10.53$

Free amino acid

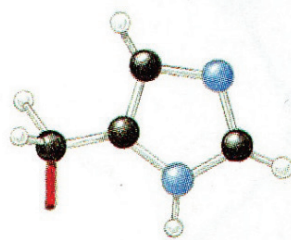
N-terminal amino group, $pK_a \sim 7.4$
C-terminal acidic group, $pK_a \sim 3.9$

In a protein

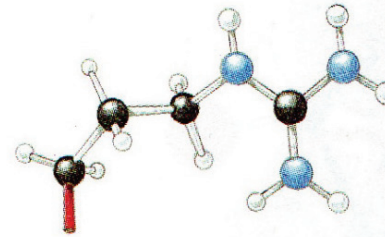
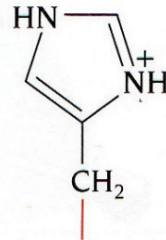
Table 1.2 Intrinsic pK_a Values of Ionizable Groups Found in Proteins

Group	Observed pK_a^a
α -Amino	6.8–8.0
α -Carboxyl	3.5–4.3
β -Carboxyl (Asp)	3.9–4.0
γ -Carboxyl (Glu)	4.3–4.5
δ -Guanido (Arg)	12.0
ϵ -Amino (Lys)	10.4–11.1
Imidazole (His)	6.0–7.0
Thiol (Cys)	9.0–9.5
Phenolic hydroxyl (Tyr)	10.0–10.3

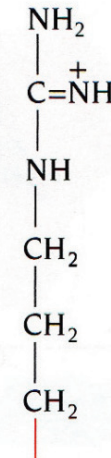
Residue properties



H His, Histidine



R Arg, Arginine



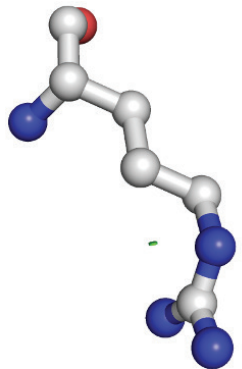
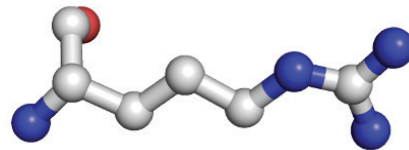
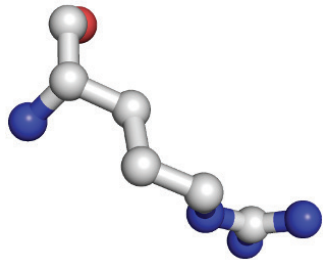
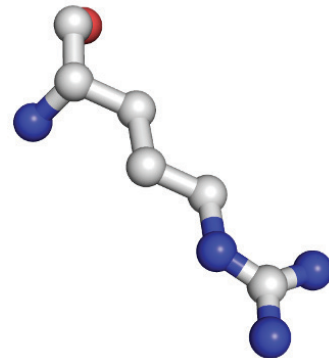
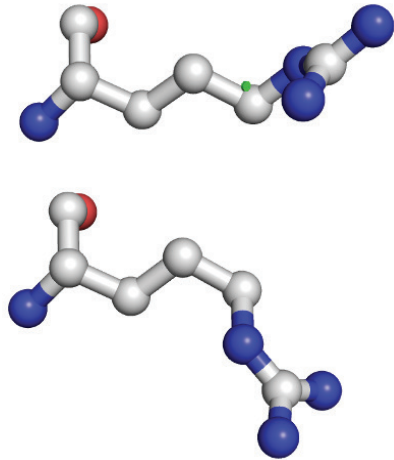
Arg is
“always”
positively
charged with
pK_a close to
12

His has pK_a close to 7 and the local environment is often tuned to to give correct acid/base chemistry. Strong base at neutral pH/Strong nucleophile. Often a catalytic residue.

Table 1.2 *Intrinsic pK_a Values of Ionizable Groups Found in Proteins*

Group	Observed pK _a ^a
α-Amino	6.8–8.0
α-Carboxyl	3.5–4.3
β-Carboxyl (Asp)	3.9–4.0
γ-Carboxyl (Glu)	4.3–4.5
δ-Guanido (Arg)	12.0
ε-Amino (Lys)	10.4–11.1
Imidazole (His)	6.0–7.0
Thiol (Cys)	9.0–9.5
Phenolic hydroxyl (Tyr)	10.0–10.3

Side chain conformations (Rotamers)



Some of *many* possibly
side chain conformations
(rotamers) for Arg

Analysis of many structures have shown that residues prefer one or a few conformations. These are called *rotamers* and are collected and distributed in *rotamer libraries*

These libraries are used in computational modeling of protein 3D structure.

Very simply put:

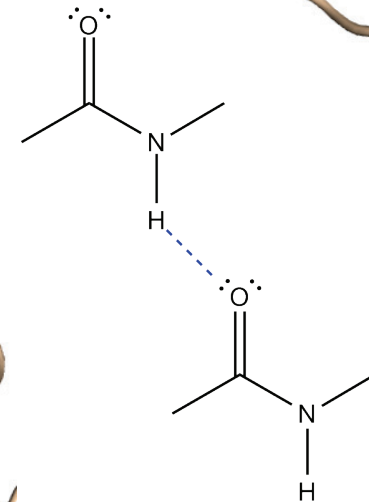
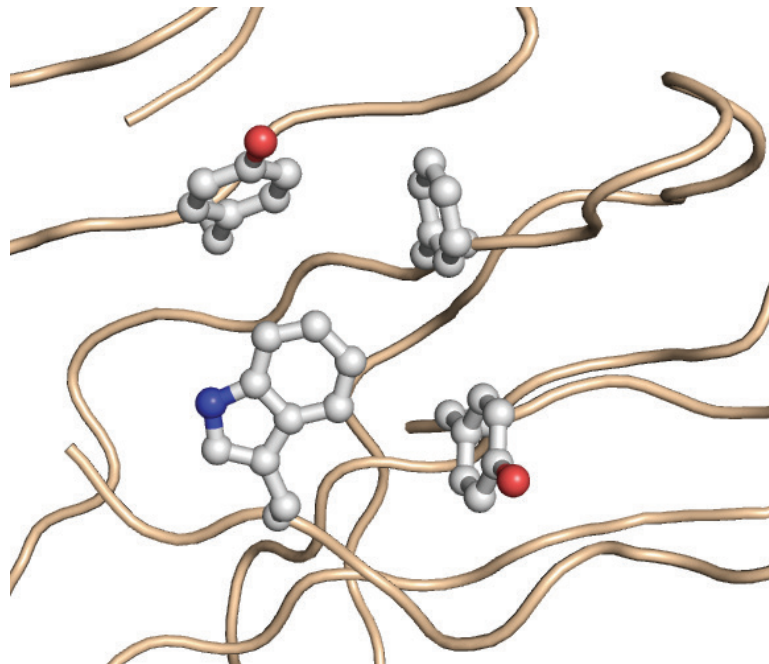
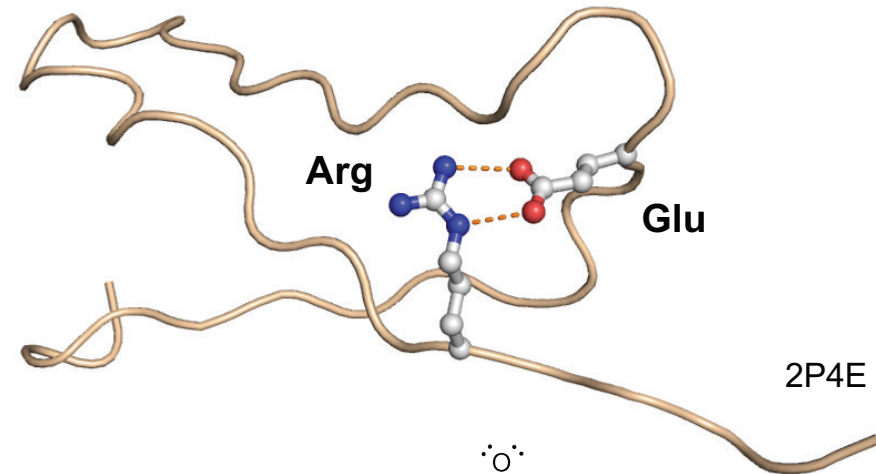
1. Determine overall 3D structure of backbone
2. Add side chains
3. Optimize side chains using conformations from rotamer libraries

Stabilizing forces

Jon K. Lærdahl,
Structural Bioinformatics

What is making proteins fold and associate into a well-defined 3D structure?

- Electrostatic interactions (salt bridges)
- Hydrogen bonds (H-bonds)
- van der Waals forces (weak)
- **IMPORTANT:** Hydrophobic interaction forces (minimizing the surface area of hydrophobic side chains exposed to solvent)

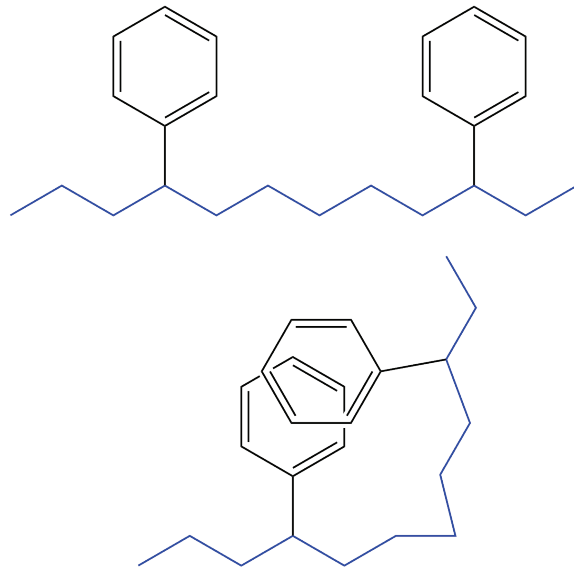
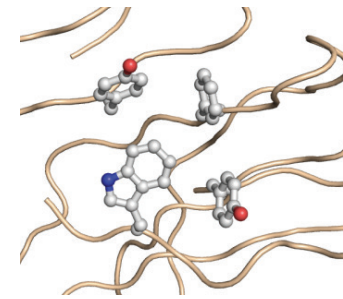


2P4E

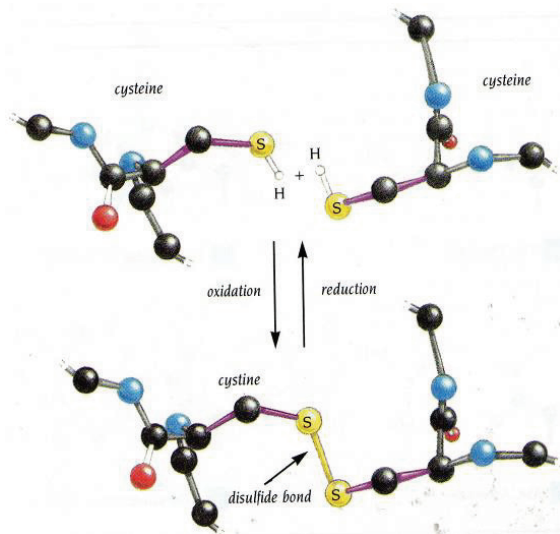
Stabilizing forces

Jon K. Lærdahl,
Structural Bioinformatics

IMPORTANT: Hydrophobic interaction forces
(minimizing the surface area of hydrophobic side
chains exposed to solvent)

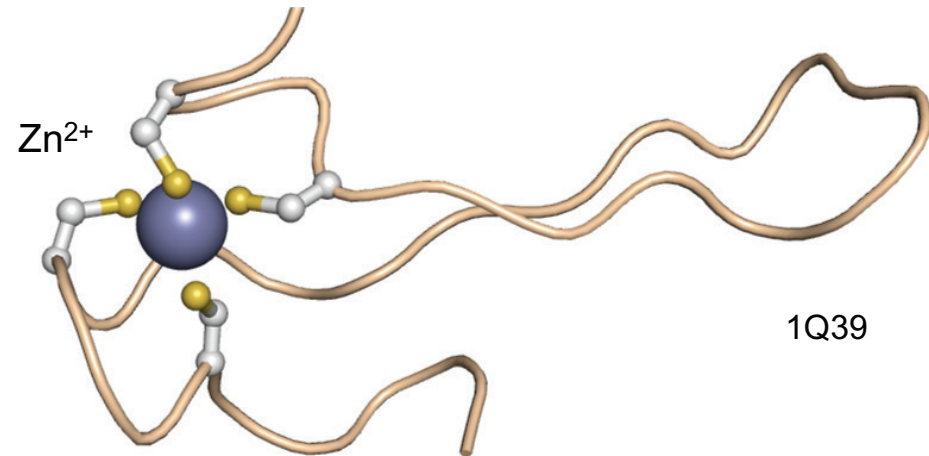


Reduced surface area
exposed to solvent (water)
for the hydrophobic side
chains



Covalent Cys-Cys
disulfide bonds

Introduction to Protein Structure,
C. Branden & J. Tooze
(Garland, New York, 1998)



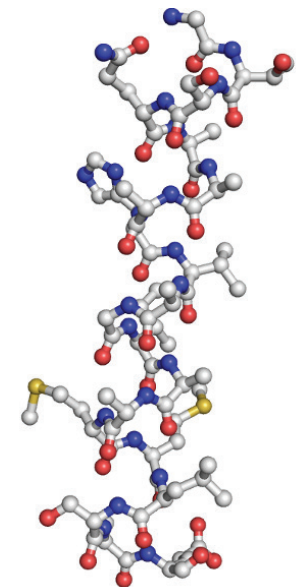
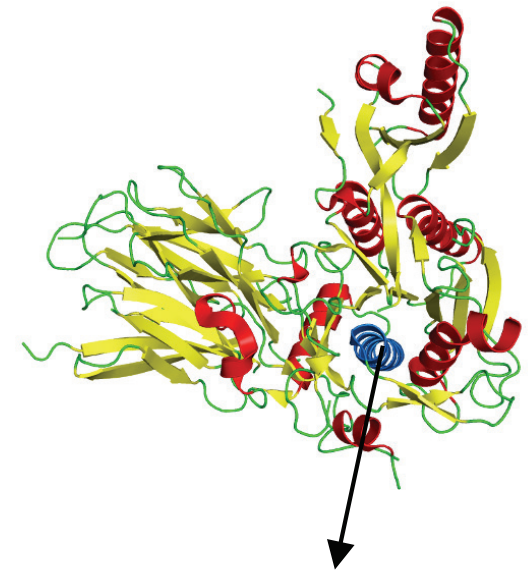
Metal ions may stabilize
the protein structure (e.g.
in zinc fingers)

Protein folding

Jon K. Lærdahl,
Structural Bioinformatics

What is making proteins fold and associate into a well-defined 3D structure?

- Proteins are often found in water and both protein-protein and protein-water interactions must be taken into account (*i.e.* interactions in folded vs. denatured state)
- *Dominant* forces responsible for tertiary structure are (believed to be) the hydrophobic interaction forces
 - Residues with hydrophobic side chains are packed in the interior of the protein
 - Charged and polar residues tend to be on the protein surface
 - Polar backbone in the protein interior is “hidden” by building secondary structure elements
- Polar residue side chains in the core must be “neutralized” by interacting with other residues, e.g. in H-bond donor-acceptor pairs
- Charged residue side chains in the core must be “neutralized” by interacting with other residues through salt bridges

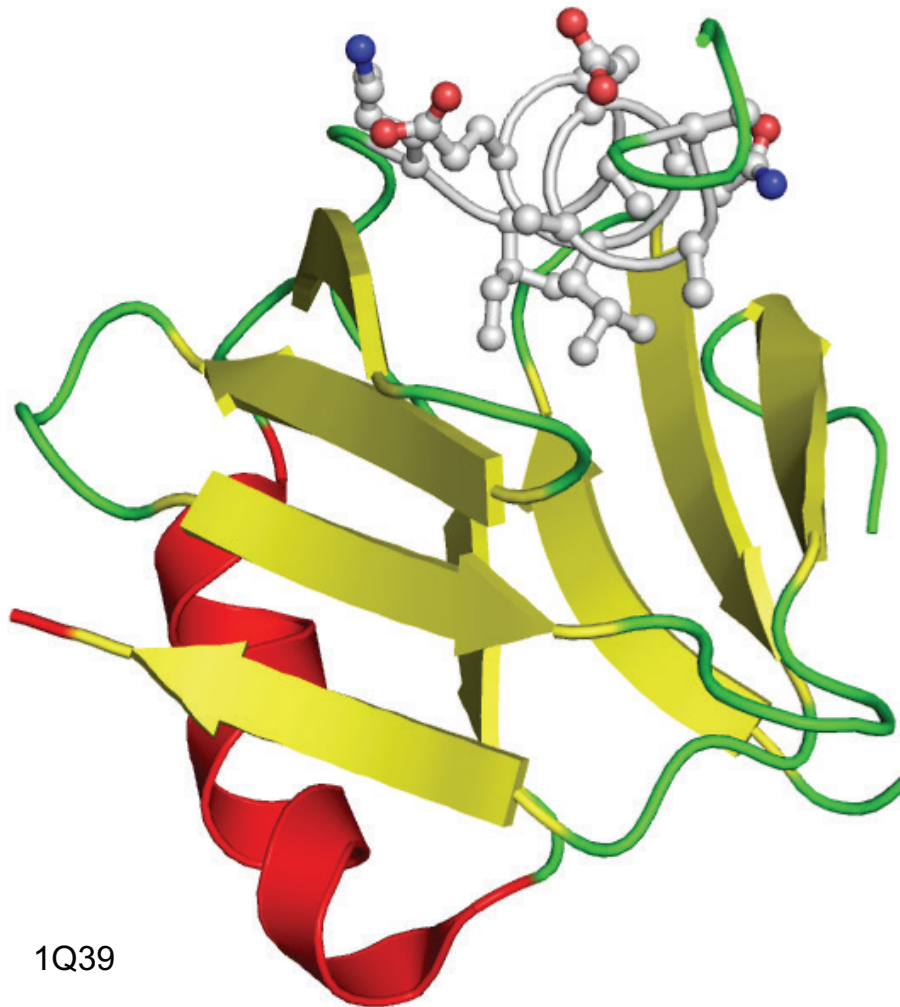


2P4E

Protein folding

Jon K. Lærdahl,
Structural Bioinformatics

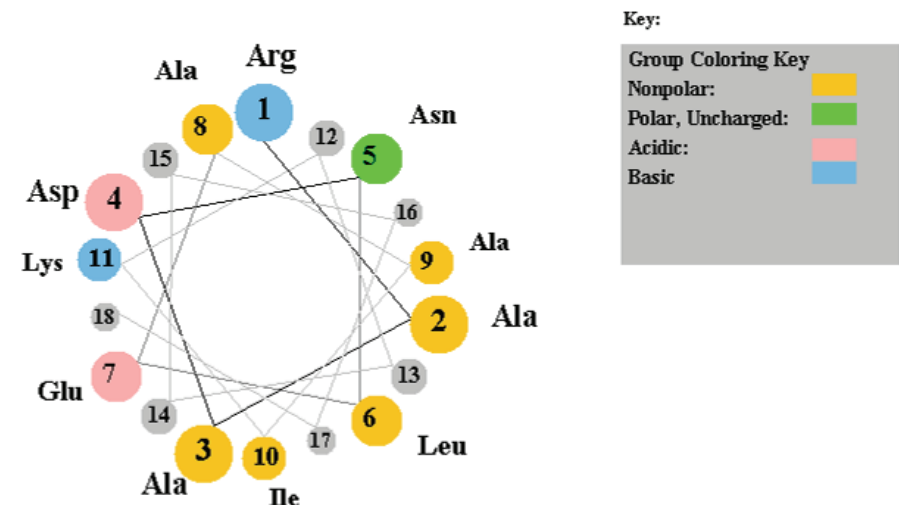
Secondary structure elements (α -helices & β -sheets) on the surfaces of proteins are often amphipathic (one hydrophilic and one hydrophobic side)



1Q39

“Pattern” of every 3-4 residues hydrophobic

Patterns can be used for predictions by computational methods, *e.g.* predict secondary structure from primary sequence



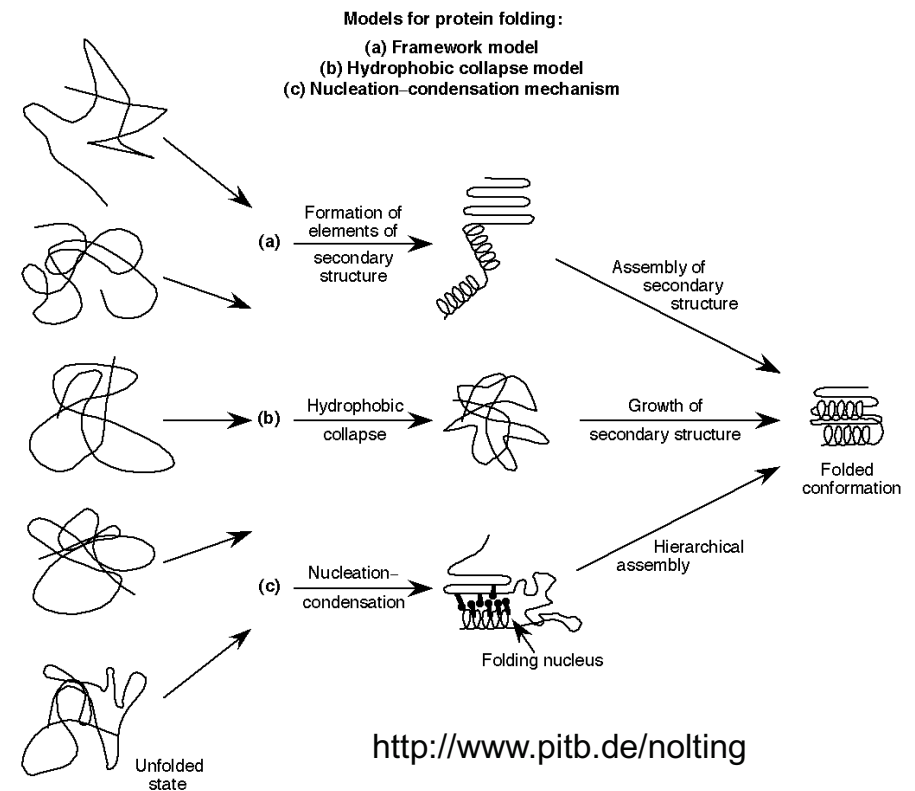
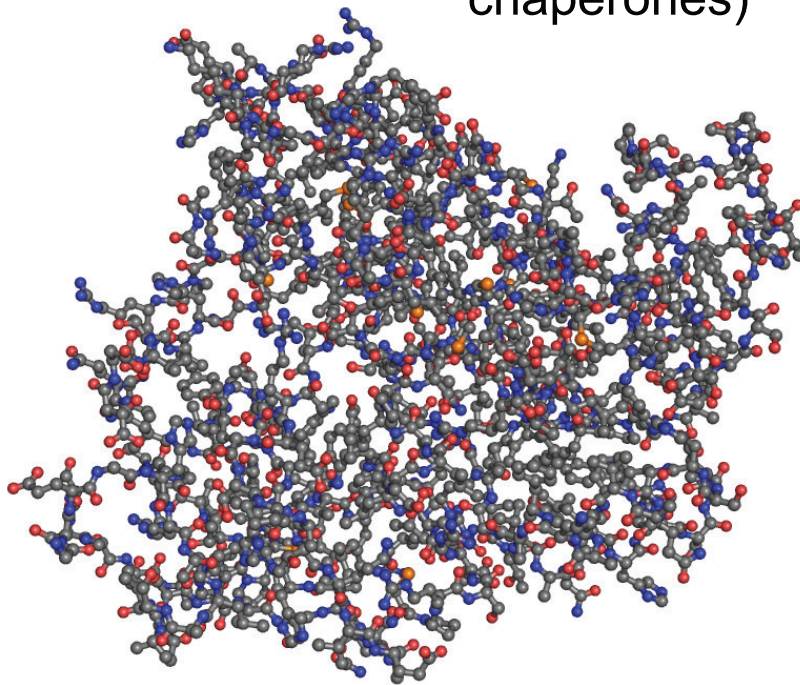
<http://cti.itc.virginia.edu/~cmg/Demo/wheel/wheelApp.html>

Protein folding

TLASTPALWASIPCRSELRLDLV
LPSEGQS



Folding is spontaneous in the cell (but often with helper molecules, chaperones)



Put very simply:

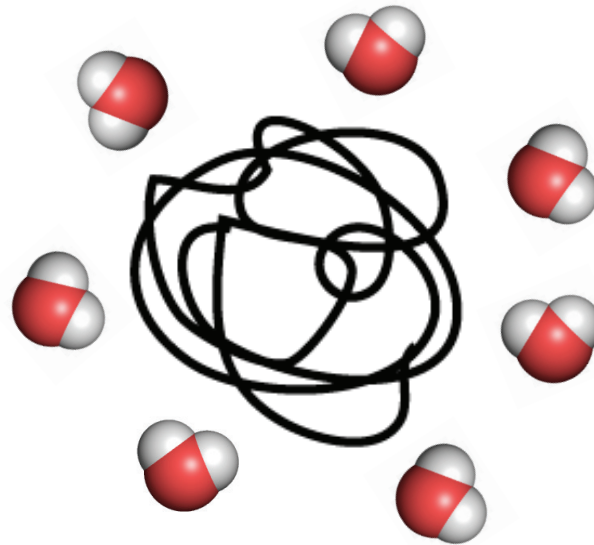
1. Secondary structure forms transiently
2. Hydrophobic collapse, formation of stable secondary structure
3. Folding completes, formation of tertiary interactions

Globular vs. membrane proteins

Jon K. Lærdahl,
Structural Bioinformatics

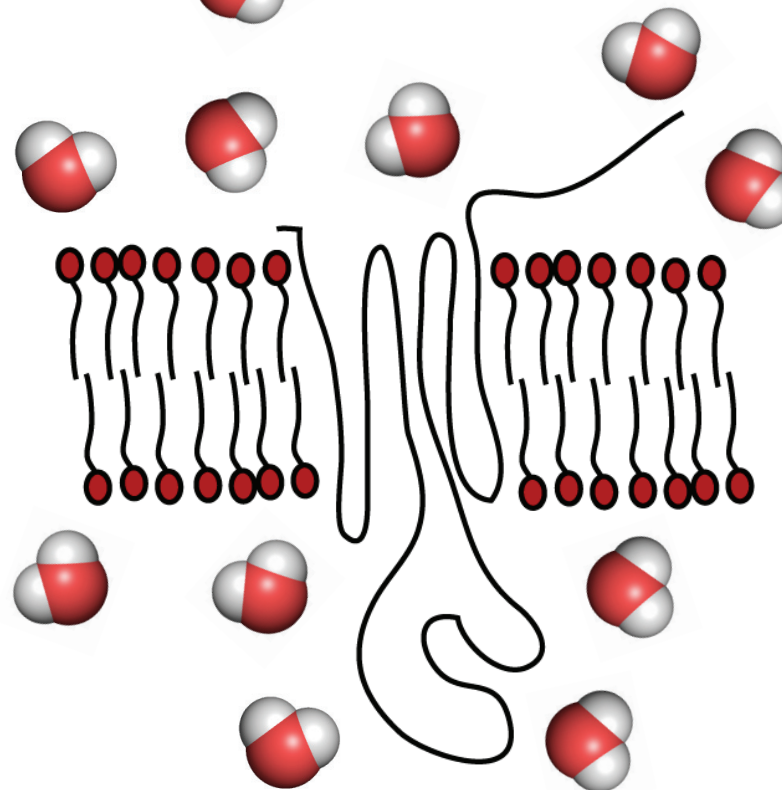
Globular proteins

- Soluble
- Surrounded by water

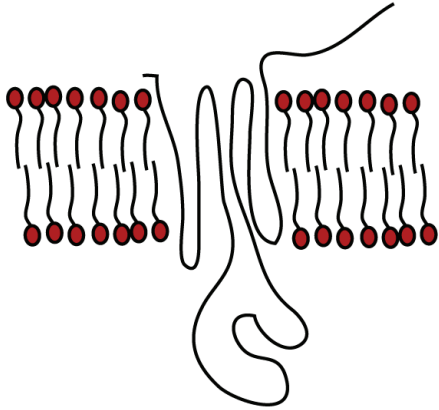


Membrane proteins

- In lipid bilayers
- Hydrophobic surface facing membrane interior

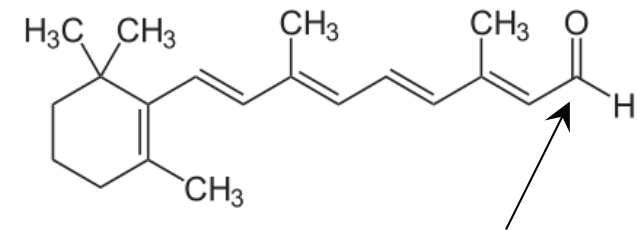


Membrane proteins

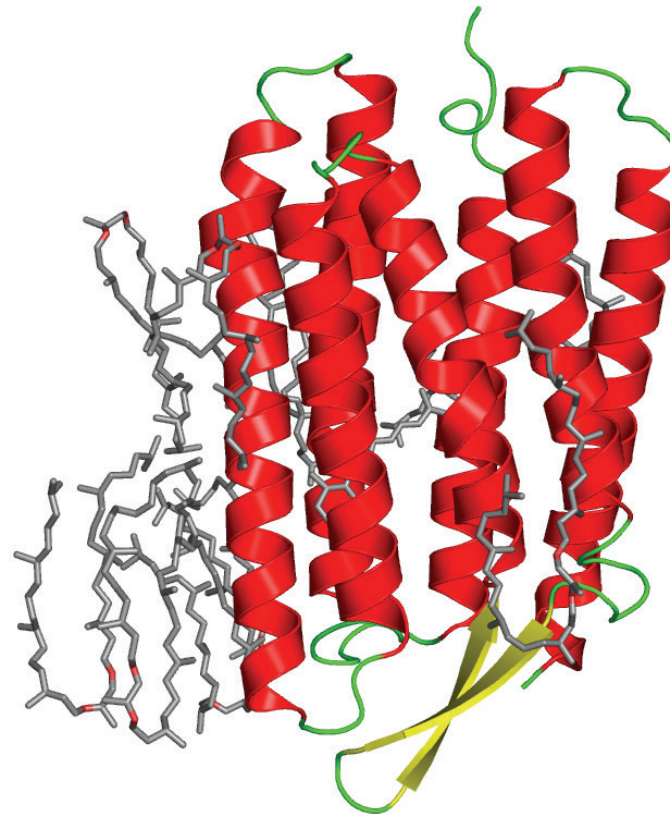


Rhodopsin (1QHJ)

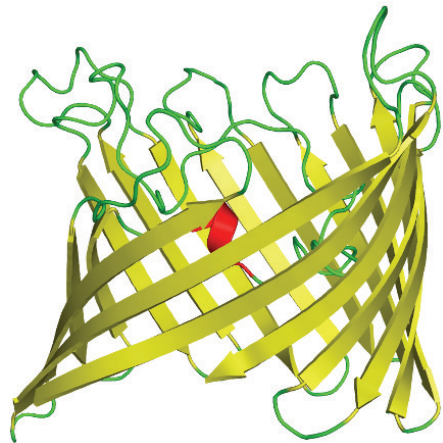
Co-factor/prosthetic group retinal:



Covalent (Schiff bond)
linkage to protein Lys
residue



Many apo-proteins need co-
factors/prosthetic groups to
become functional



Beta-barrel porin (1PRN)

